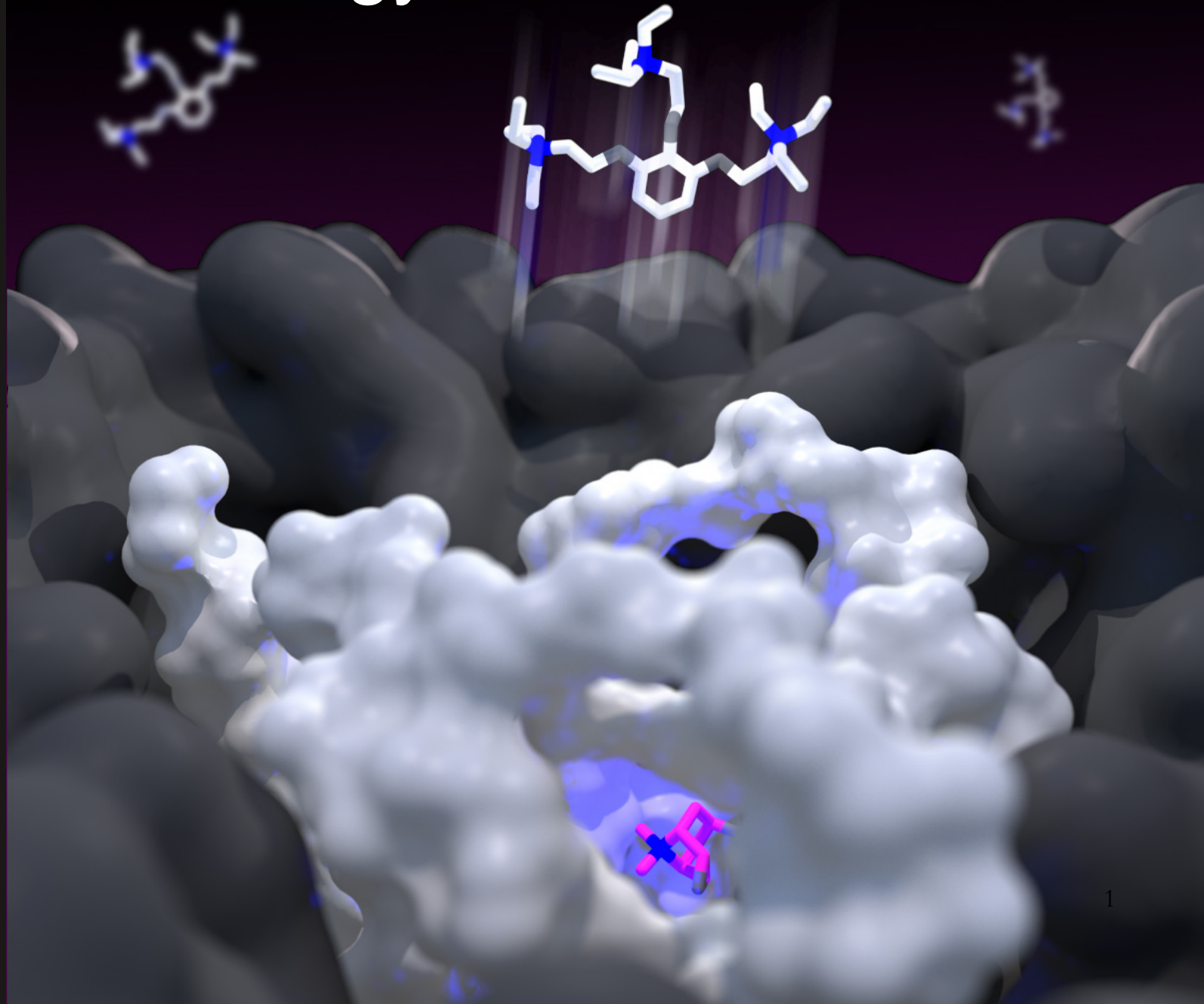


CS/CME/Biophys/BMI 371

Computational biology in four dimensions

Ron Dror

Jan. 9, 2017



*Image credit:
Sam Hertig*

Outline for lecture 1 (course overview)

- Course format and objectives
- What is structure?
 - Structure (and dynamics) at multiple spatial scales
- Why is structure important?
- Overview of course topics
 - Atomic-level modeling of biomolecules
 - Structures of macromolecular complexes
 - Cellular-level organization
- Course logistics
- Guidelines for presentations and critiques
- Immediate next steps

Course format and objectives

Focus is on presentation, discussion, and critique of cutting-edge research

- The majority of class time will be spent on presentations and discussion of important recent research papers
- Most presentations will be by students
 - Each student will present on a paper of interest to them
 - Professor & TAs will meet with students to review presentations
 - Professor will introduce topics in class
 - Professor and guest speaker will give initial presentations
- Professor will facilitate discussion. Students are strongly encouraged to share comments, questions

Course requirements

- One presentation
 - 20-30 minutes each
- Three written critiques of papers being presented
- Read the remaining papers, attend class, and participate in discussions

What do I want students to learn from this course?

1. Gain exposure to cutting-edge computational research in structural biology, broadly defined (a rapidly growing, interdisciplinary area).
2. Learn to critique and evaluate research, and practice critical reading of research papers.
3. Refine the skill of presenting deep technical material to a non-expert audience.

The latter two are broadly important skills. They're especially important to practice if they make you a bit nervous.

Relationship to CS/CME/BioE/Biophys/BMI 279

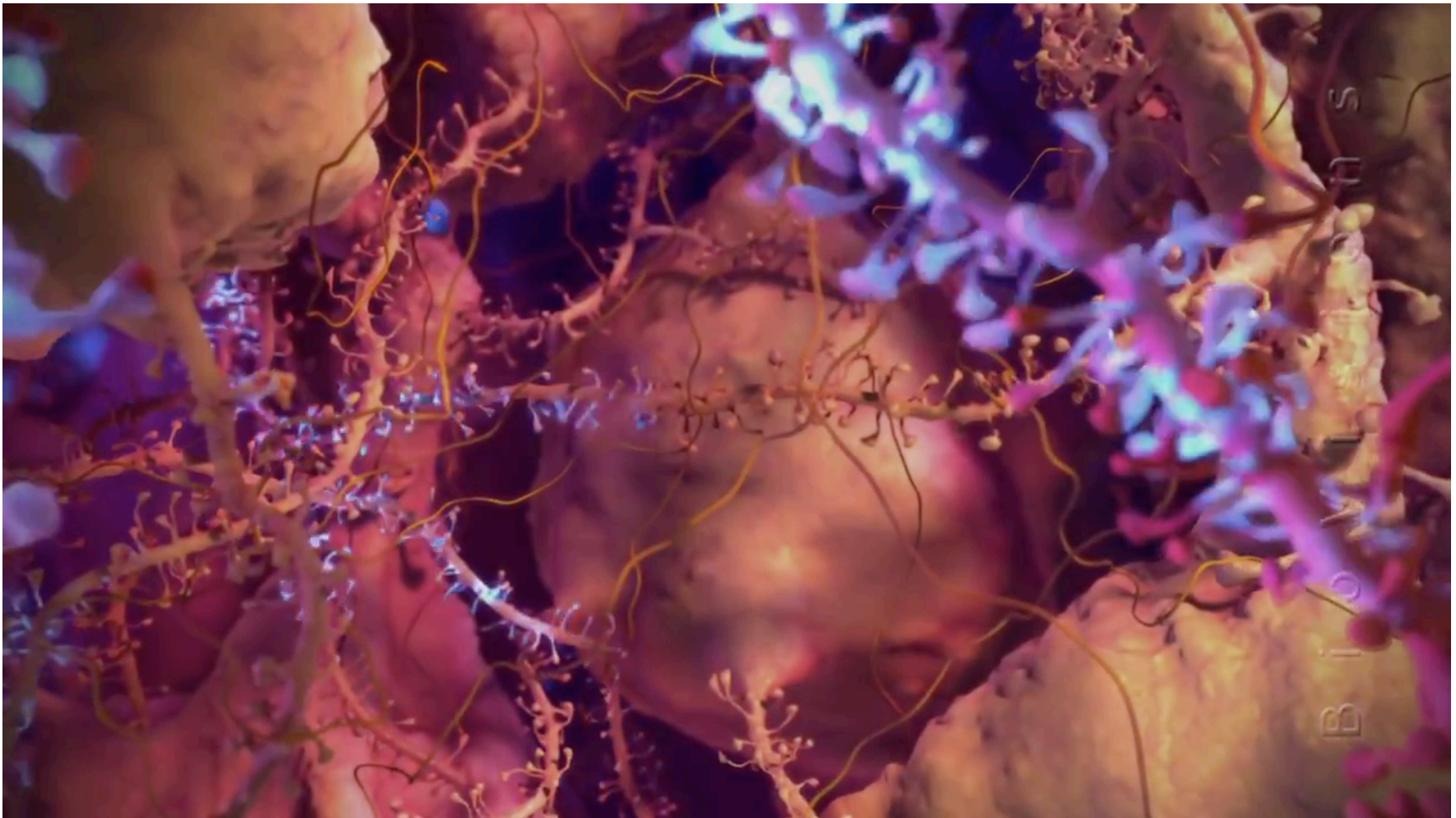
- 371 and 279 cover the same general field
- 279 is a traditional lecture/homework course covering basics of the field
- 371 focuses on current research topics
- Neither is a prerequisite for the other

What is structure?

In daily life, we use machines
with functional *structure* and *moving parts*



Cells and biomolecules (e.g., proteins) are also machines whose function depends on structure and moving parts

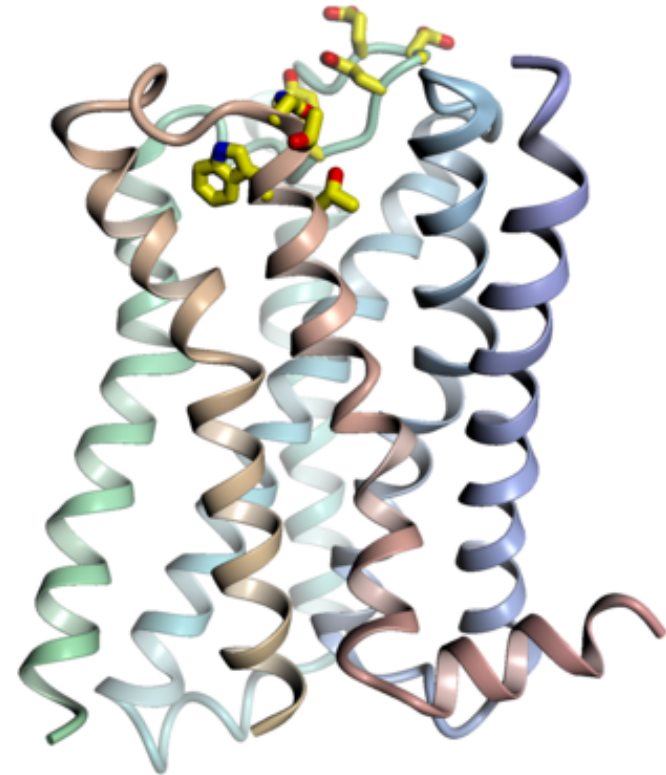
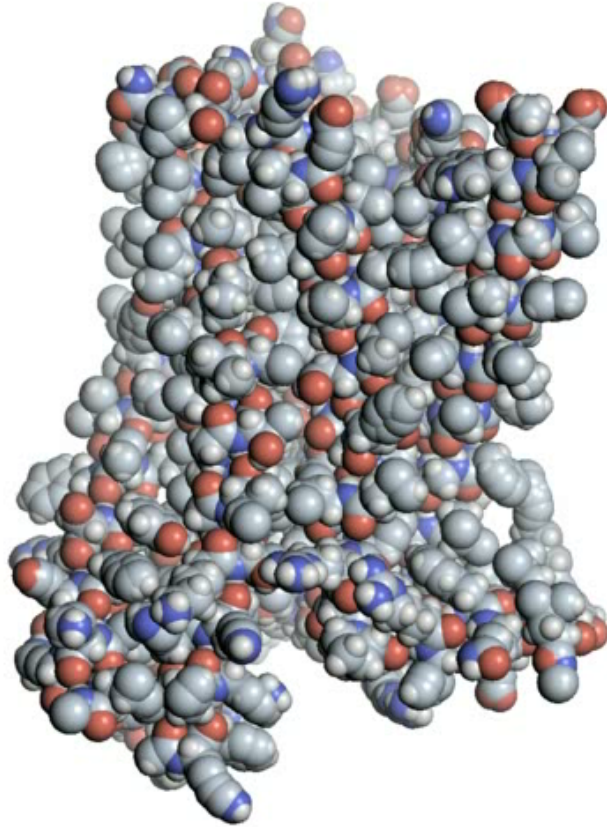


From *Inner Life of the Cell* | *Protein Packing*, XVIVO and Biovisions @ Harvard

What is structure?

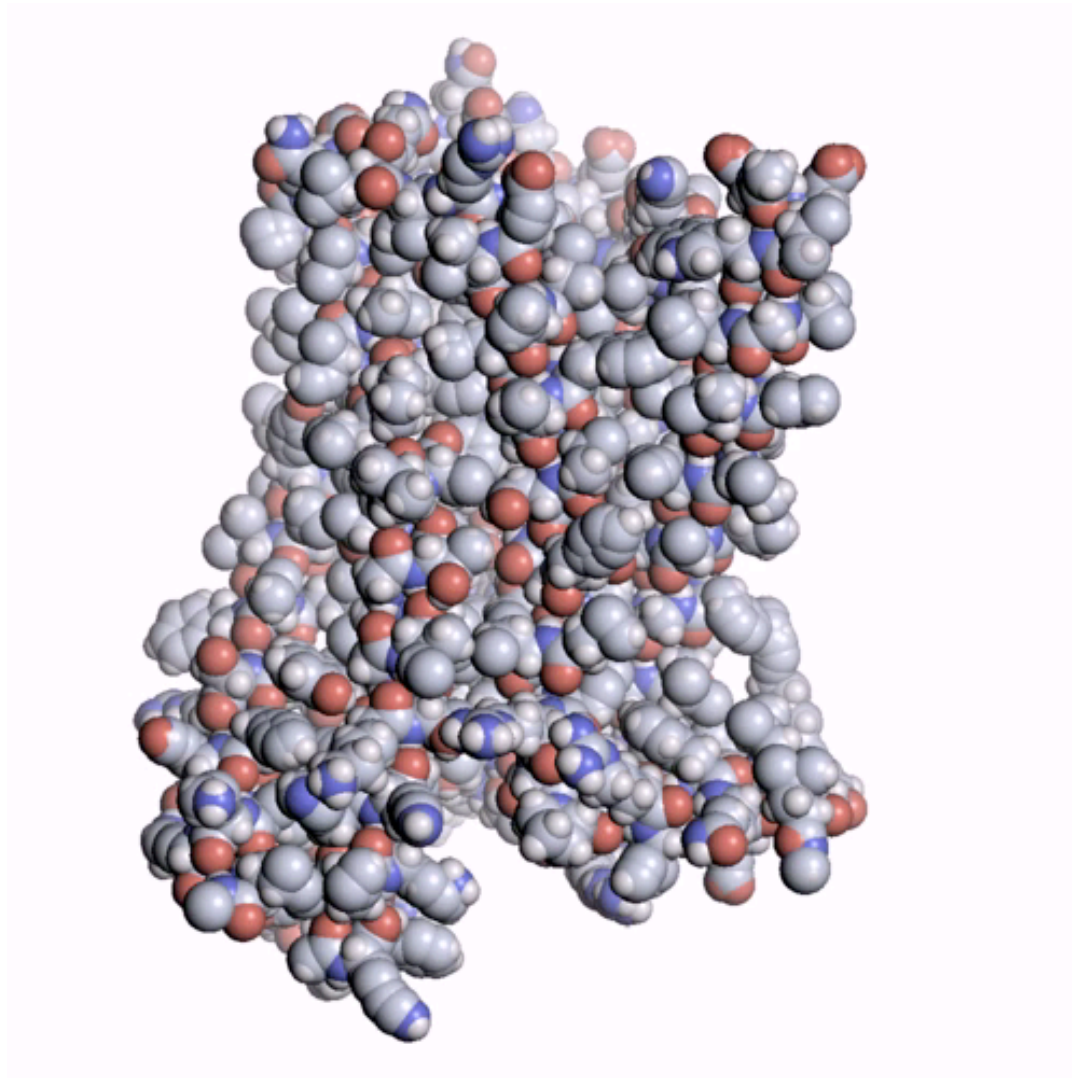
**Structure (and dynamics)
at multiple spatial scales**

Protein structure



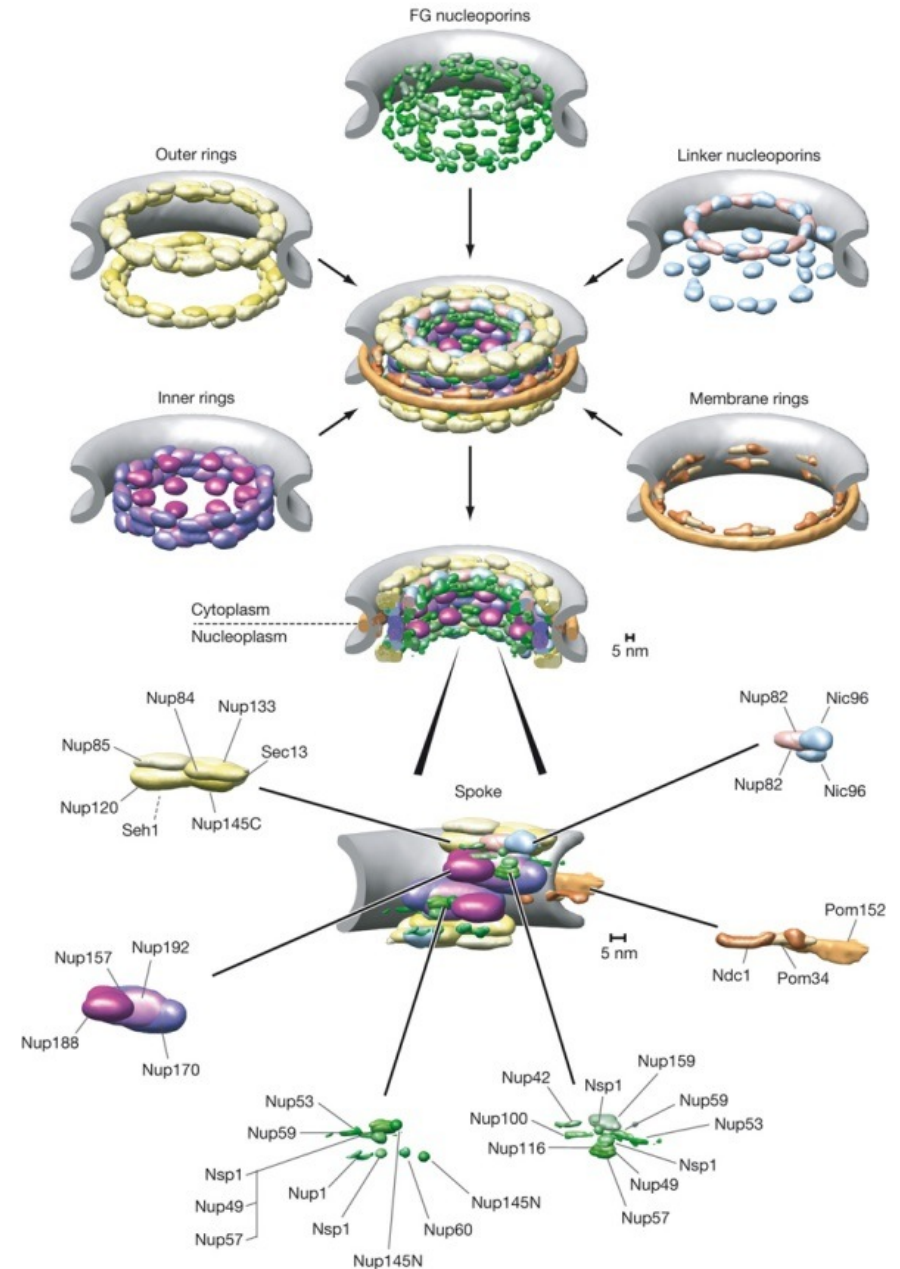
An adrenaline receptor
(the β_2 adrenergic receptor)

Protein dynamics



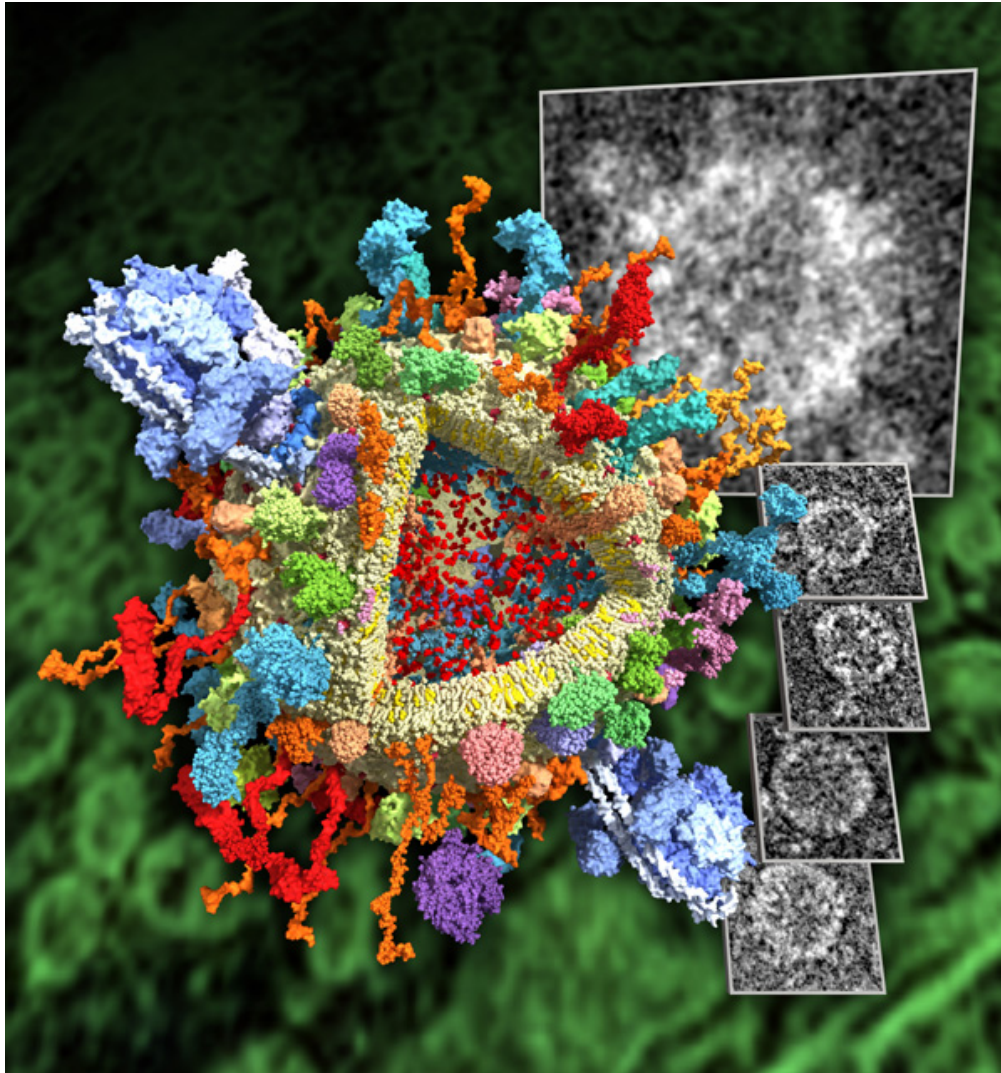
β_2 adrenergic receptor

Proteins (and other molecules) often come together to form *macromolecular complexes*



Nuclear Pore Complex
Alber et al., *Nature* 2007

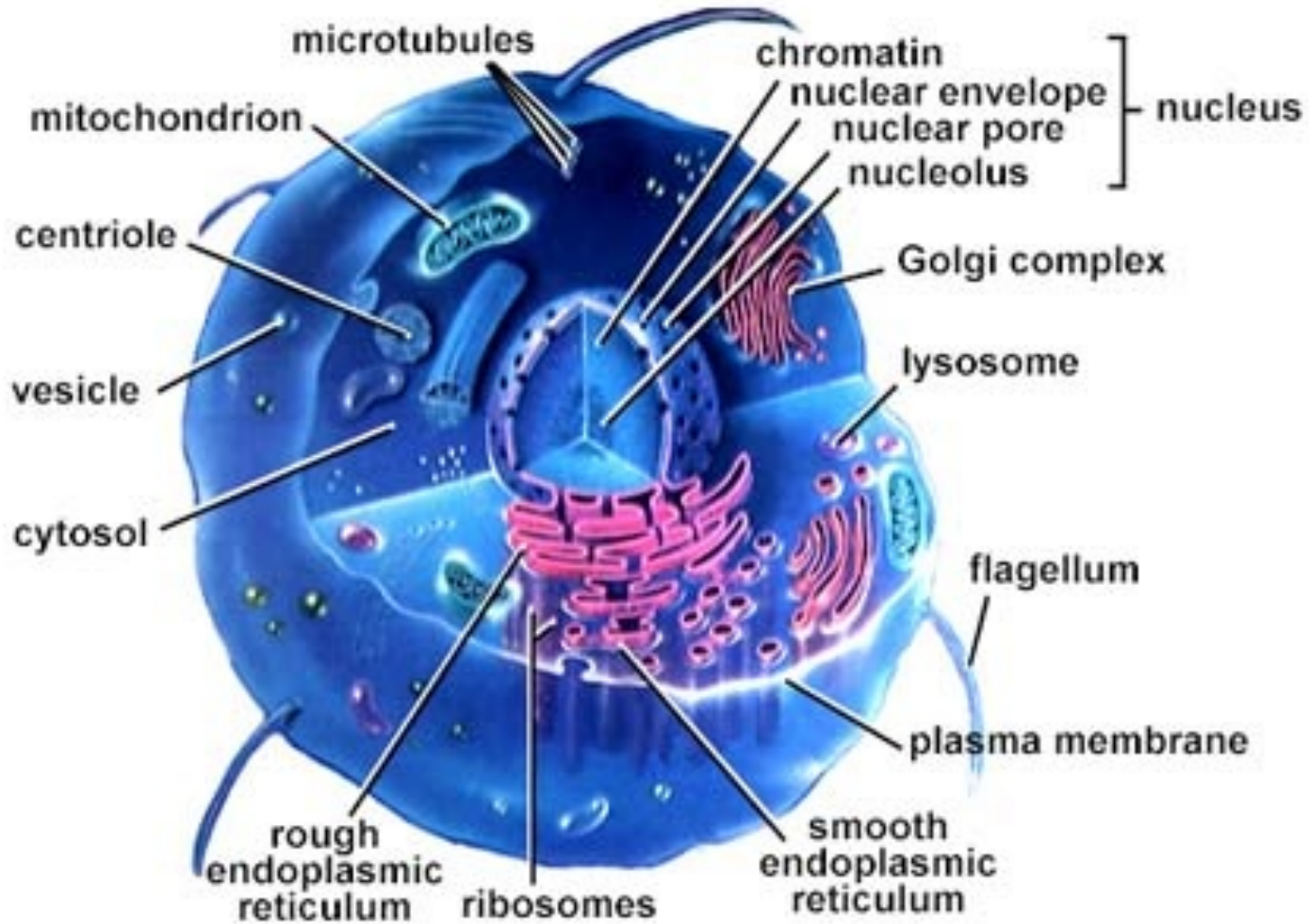
These come together to form organelles



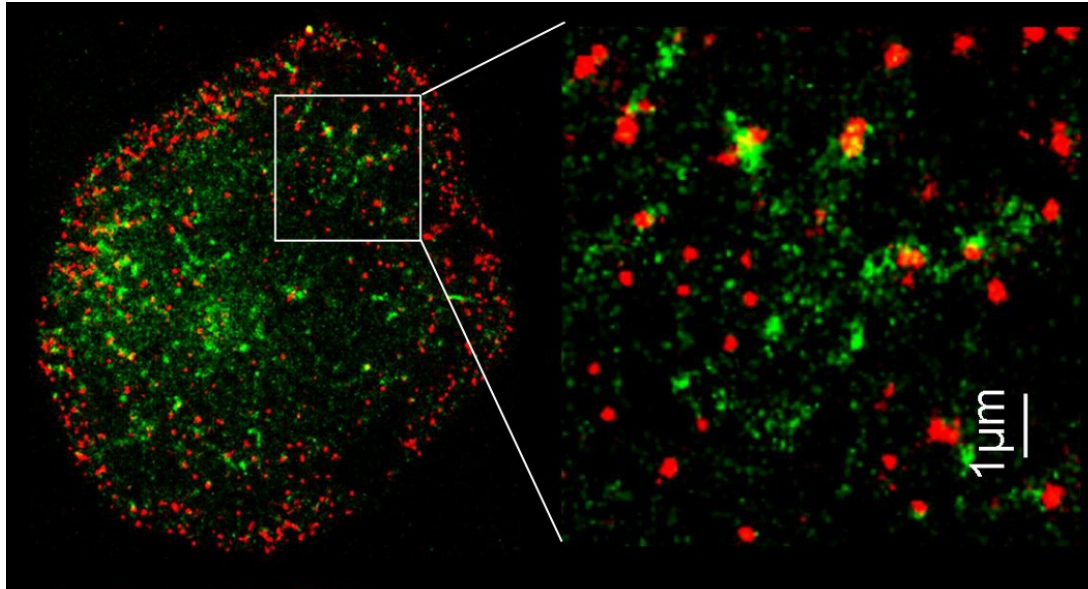
Synaptic vesicle

<http://www.mpibpc.mpg.de/9547480/vesicle600.jpg>

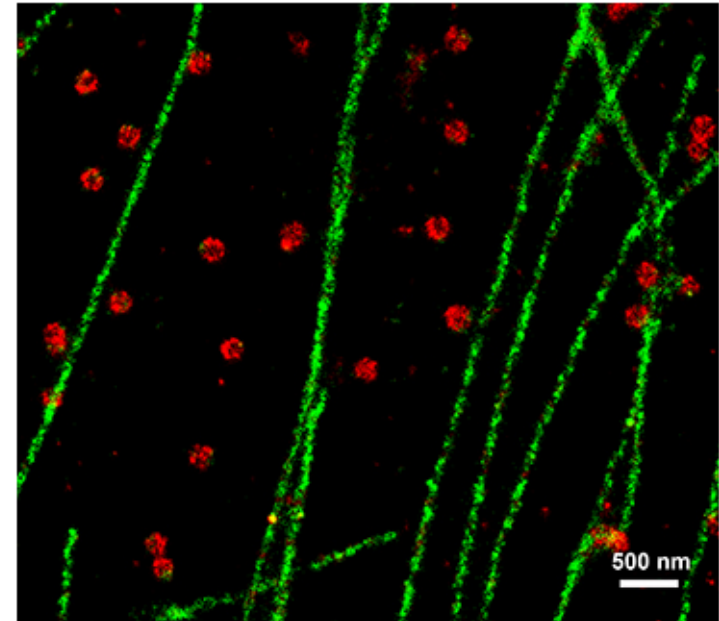
and cells



Intracellular structure



Chih-Jung Hsu, Janis Burkhardt and Tobias Baumgart

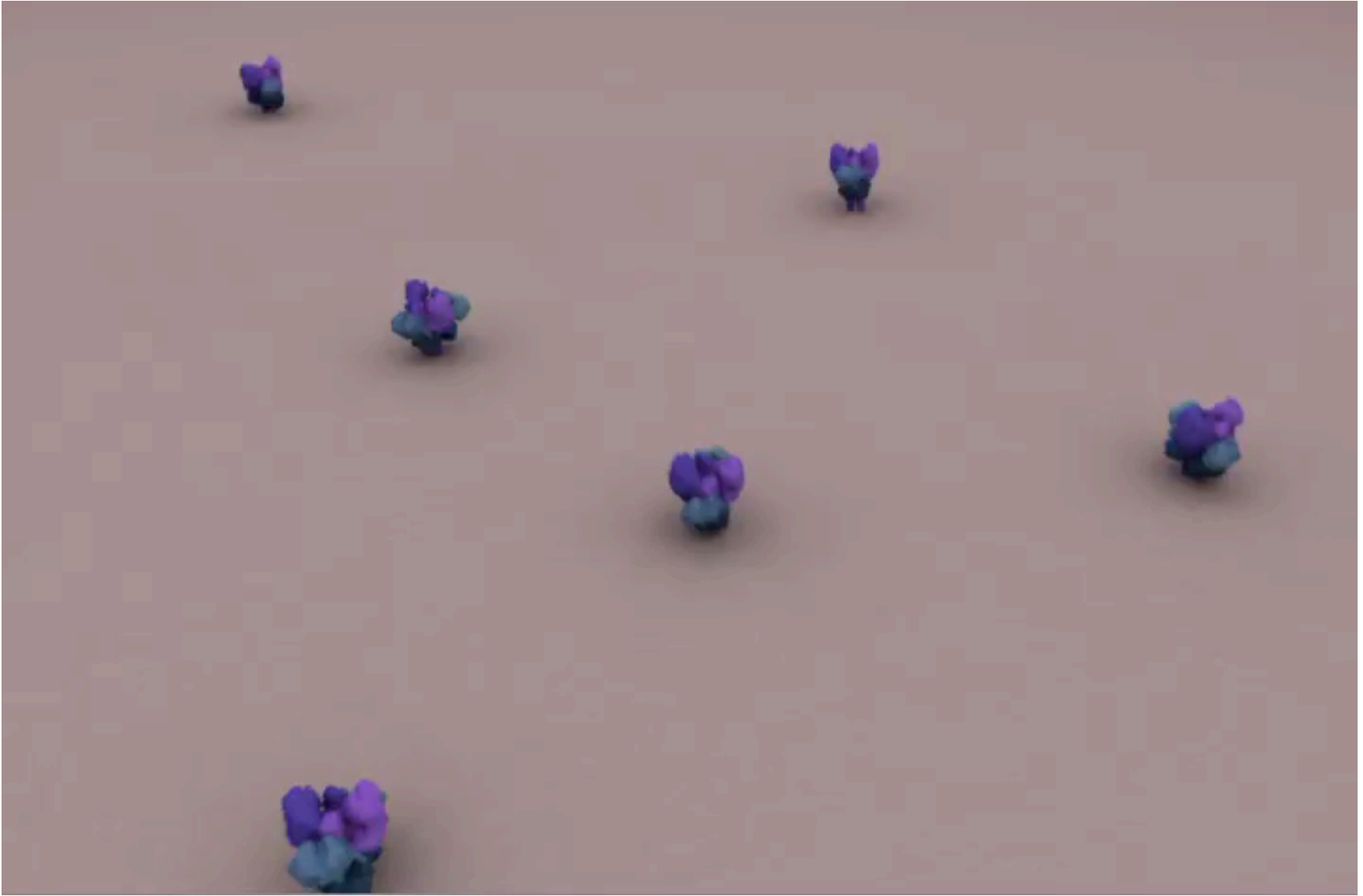


[http://www.nikoninstruments.com/Products/Microscope-Systems/Inverted-Microscopes/N-STORM-Super-Resolution/\(gallery\)](http://www.nikoninstruments.com/Products/Microscope-Systems/Inverted-Microscopes/N-STORM-Super-Resolution/(gallery)); Zhuang group



David Goodsell

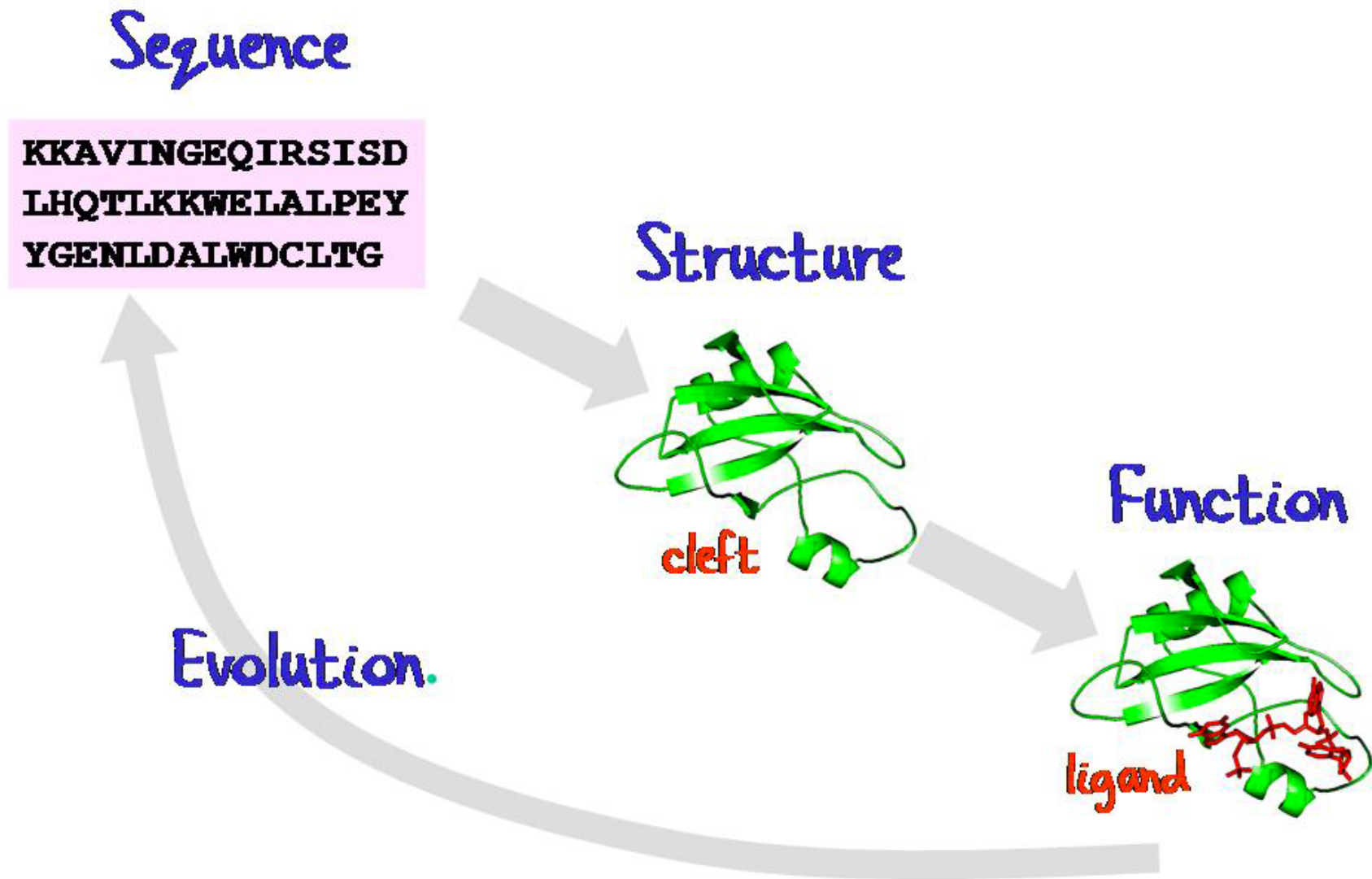
Intracellular dynamics (artist's rendition)



Janet Iwasa and Tomas Kirchhausen₁₈

Why is structure important?

The cycle of life



From Michael Levitt

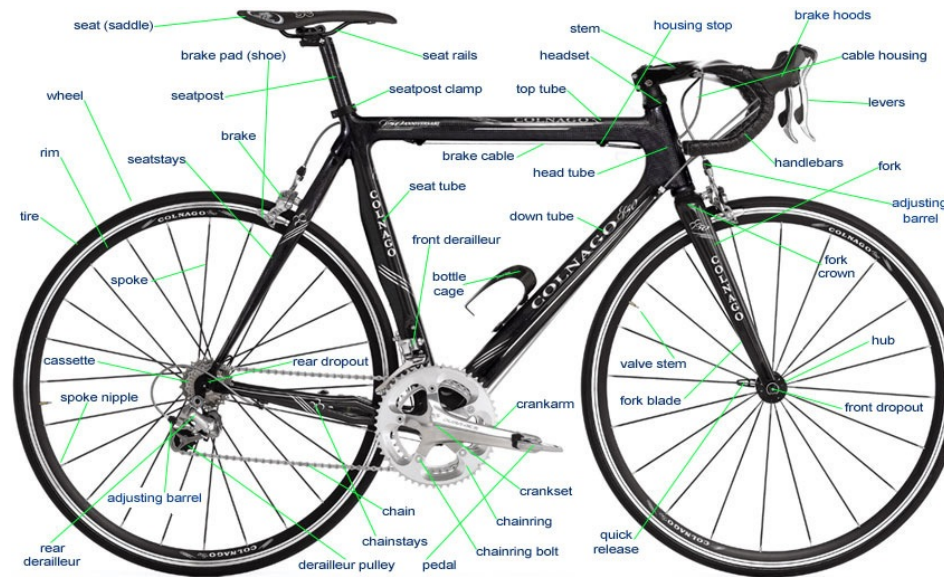
Genomics is a great start

Track Bike – DL 175

REF. NO.	IBM NO.	DESCRIPTION
1	156011	Track Frame 21", 22", 23", 24", Team Red
2	157040	Fork for 21" Frame
2	157039	Fork for 22" Frame
2	157038	Fork for 23" Frame
2	157037	Fork for 24" Frame
3	191202	Handlebar TTT Competition Track Alloy 15/16"
4		Handlebar Stem, TTT, Specify extension
5	191278	Expander Bolt
6	191272	Clamp Bolt
7	145841	Headset Complete 1 x 24 BSC
8	145842	Ball Bearings
9	190420	175 Raleigh Pistard Seta Tubular Prestavalve 27"
10	190233	Rim, 27" AVA Competition (36H) Alloy Prestavalve
11	145973	Hub, Large Flange Campagnolo Pista Track Alloy (pairs)
12	190014	Spokes, 11 5/8"
13	145837	Sleeve
14	145636	Ball Bearings
15	145170	Bottom Bracket Axle
16	145838	Cone for Sleeve
17	146473	L.H. Adjustable Cup
18	145833	Lockring
19	145239	Straps for Toe Clips
20	145834	Fixing Bolt
21	145835	Fixing Washer
22	145822	Dustcap
23	145823	R.H. and L.H. Crankset with Chainwheel
24	146472	Fixed Cup
25	145235	Toe Clips, Christophe, Chrome (Medium)
26	145684	Pedals, Extra Light, Pairs
27	123021	Chain
28	145980	Seat Post
29		Seat Post Bolt and Nut
30	167002	Saddle, Brooks
31	145933	Track Sprocket, Specify 12, 13, 14, 15, or 16 T.

- But a parts list is not enough to understand how a bicycle works

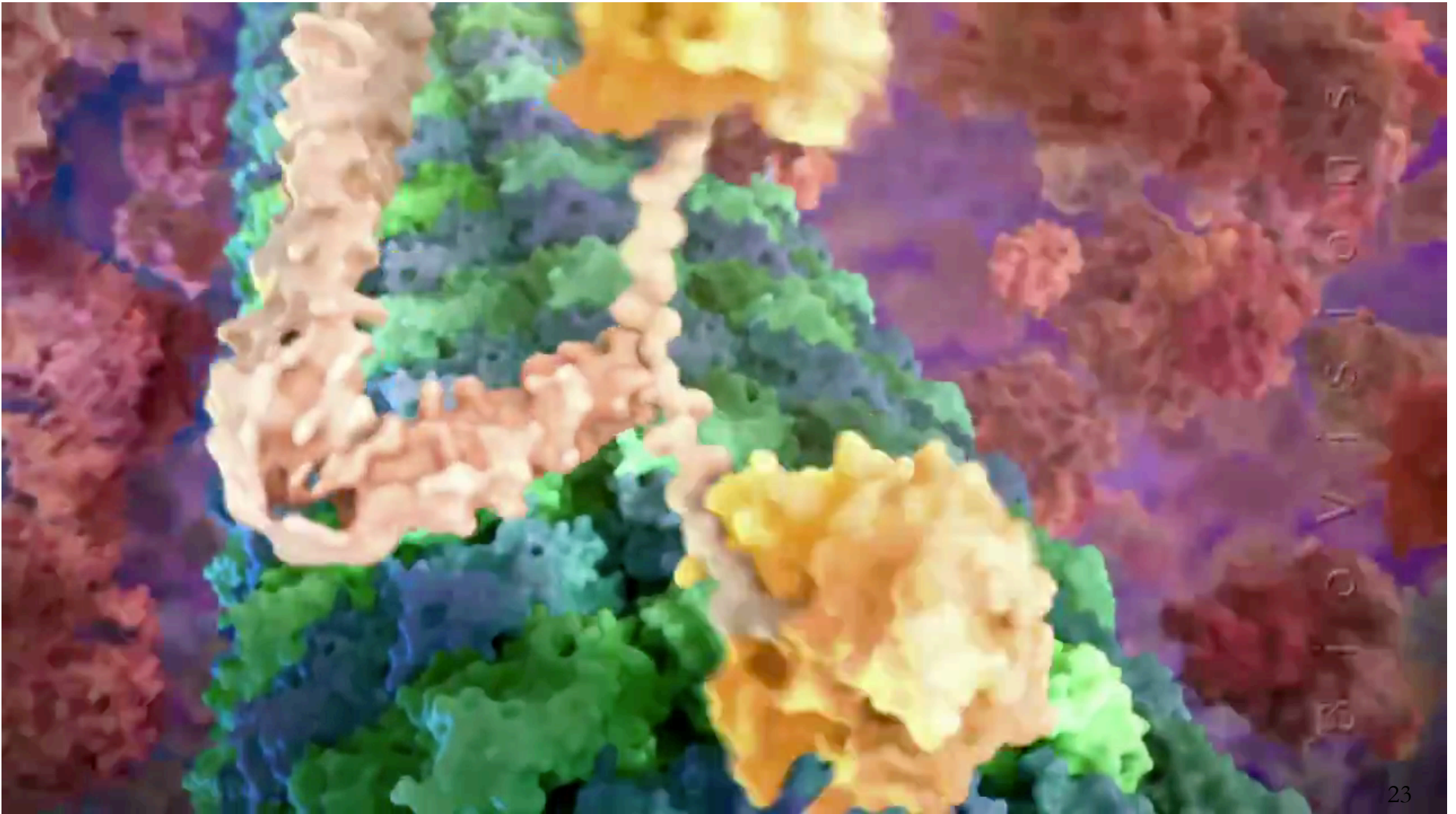
... but not the end



- We want the full spatiotemporal picture, and an ability to control it
- Broad applications, including drug design, medical diagnostics, chemical manufacturing, and energy

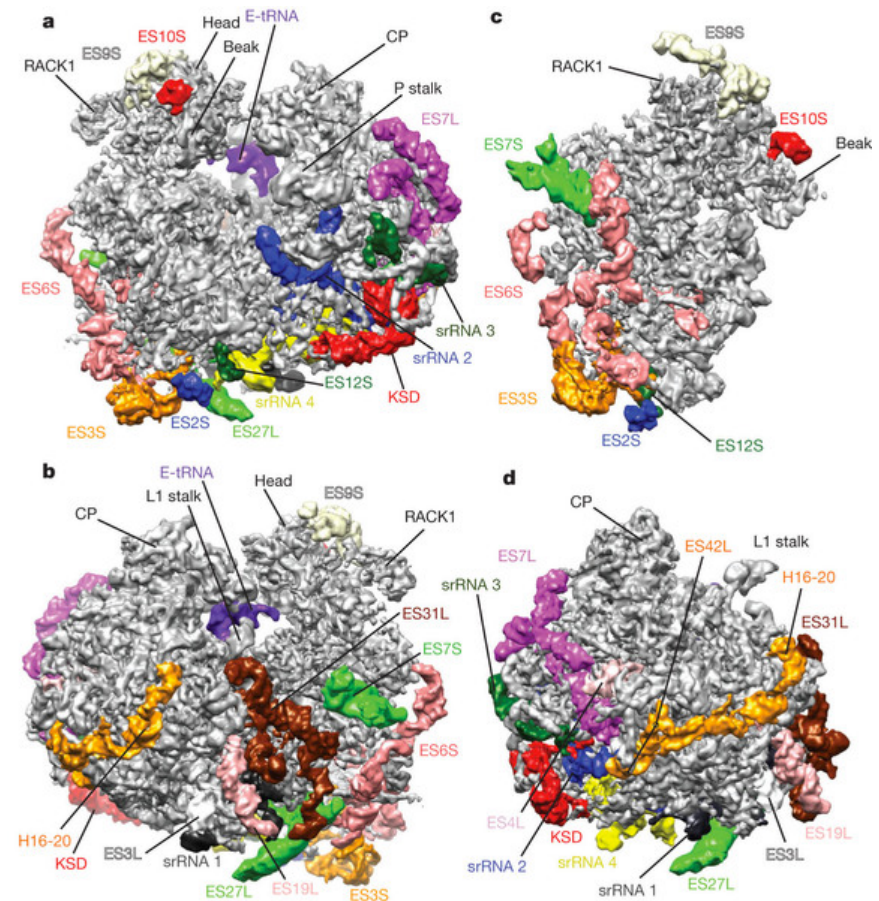
Structure determines function

- Example: Motor protein (walks along microtubules, dragging load)



Structure determines function

- Example: Ribosome
 - Complex of many proteins and RNAs that together makes new proteins (by reading the genetic code and combining amino acids)

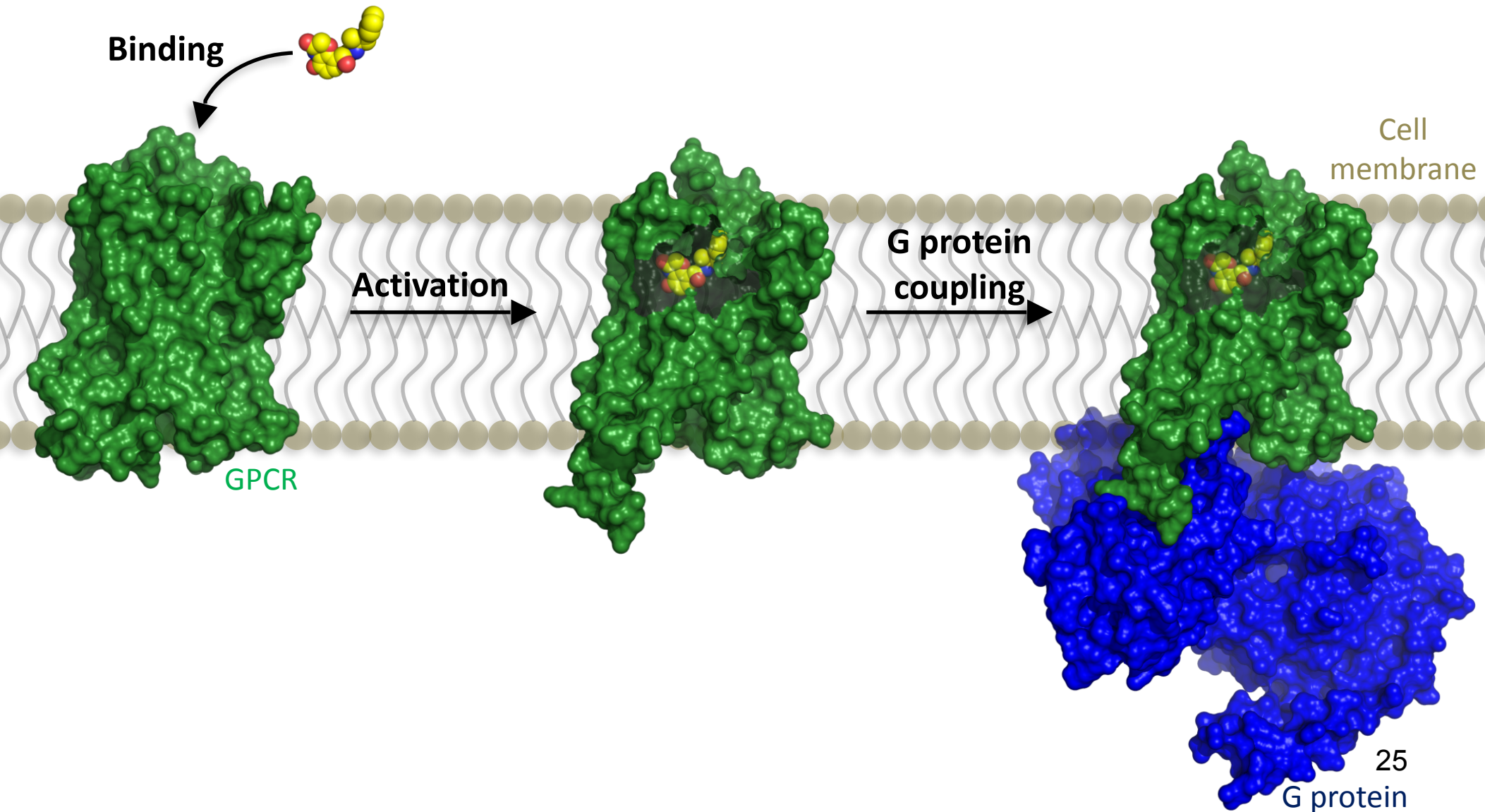


From *Inner Life of the Cell*, XVIVO and Biovisions @ Harvard

Hashem et al., Nature 494:385-9, 2013

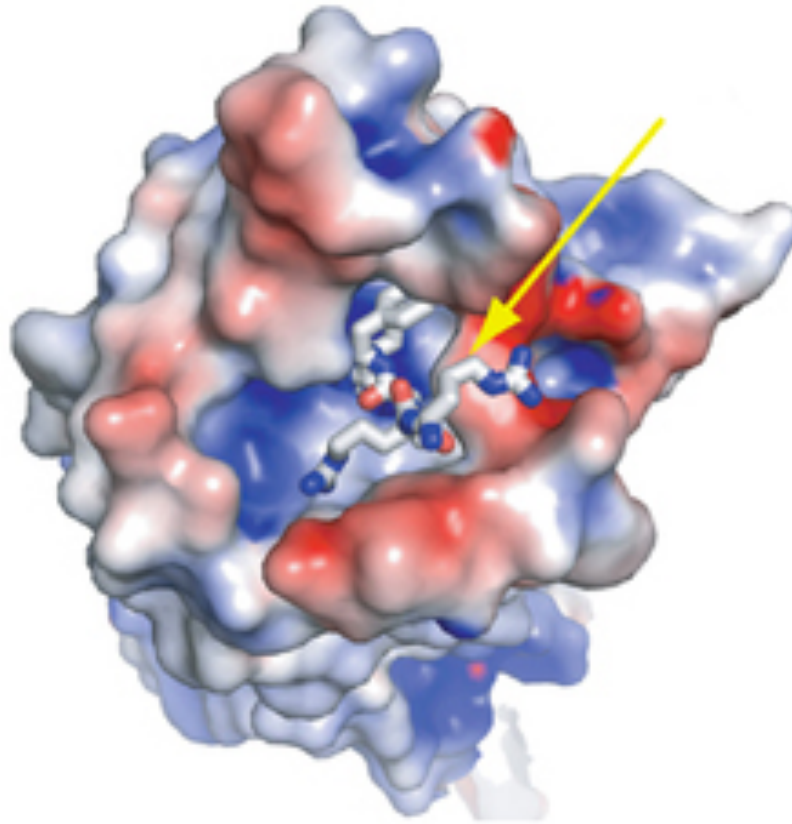
Structure determines function

- Example: G protein-coupled receptors (GPCRs)
 - Largest class of human drug targets
 - Function: allow the cell to sense and respond to molecules outside it



Structure-based drug design

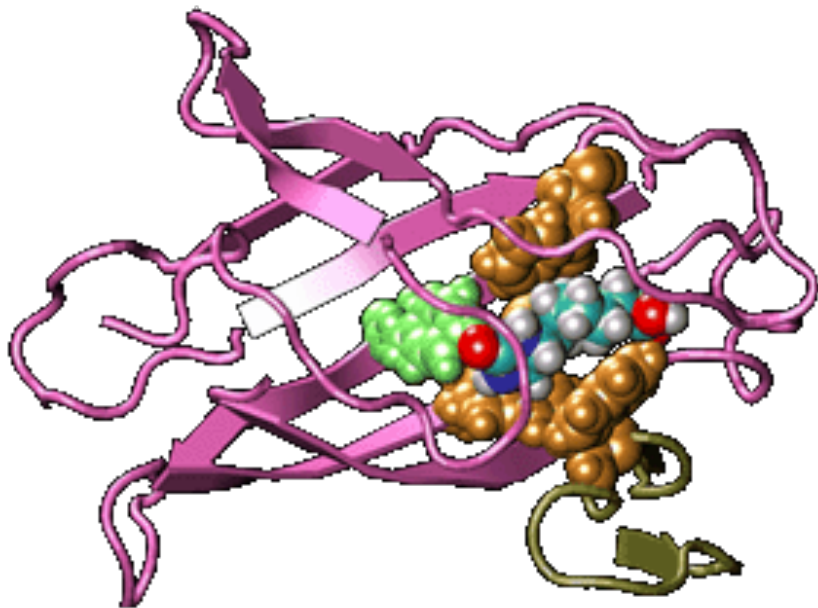
- Almost all drugs act by binding to proteins and altering their function
- Using knowledge of structures, we can design drugs that bind more tightly or more selectively, bind in different positions, alter behavior of protein in different ways, etc.



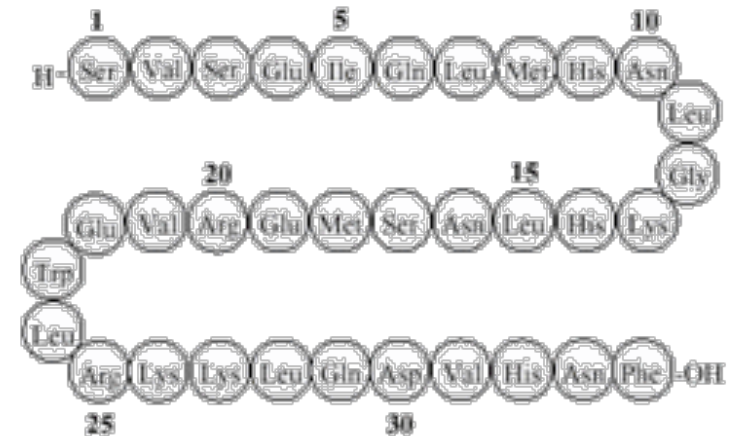
http://www.nih.gov/researchmatters/october2012/images/structure_l.jpg

Designing new biomolecular machines

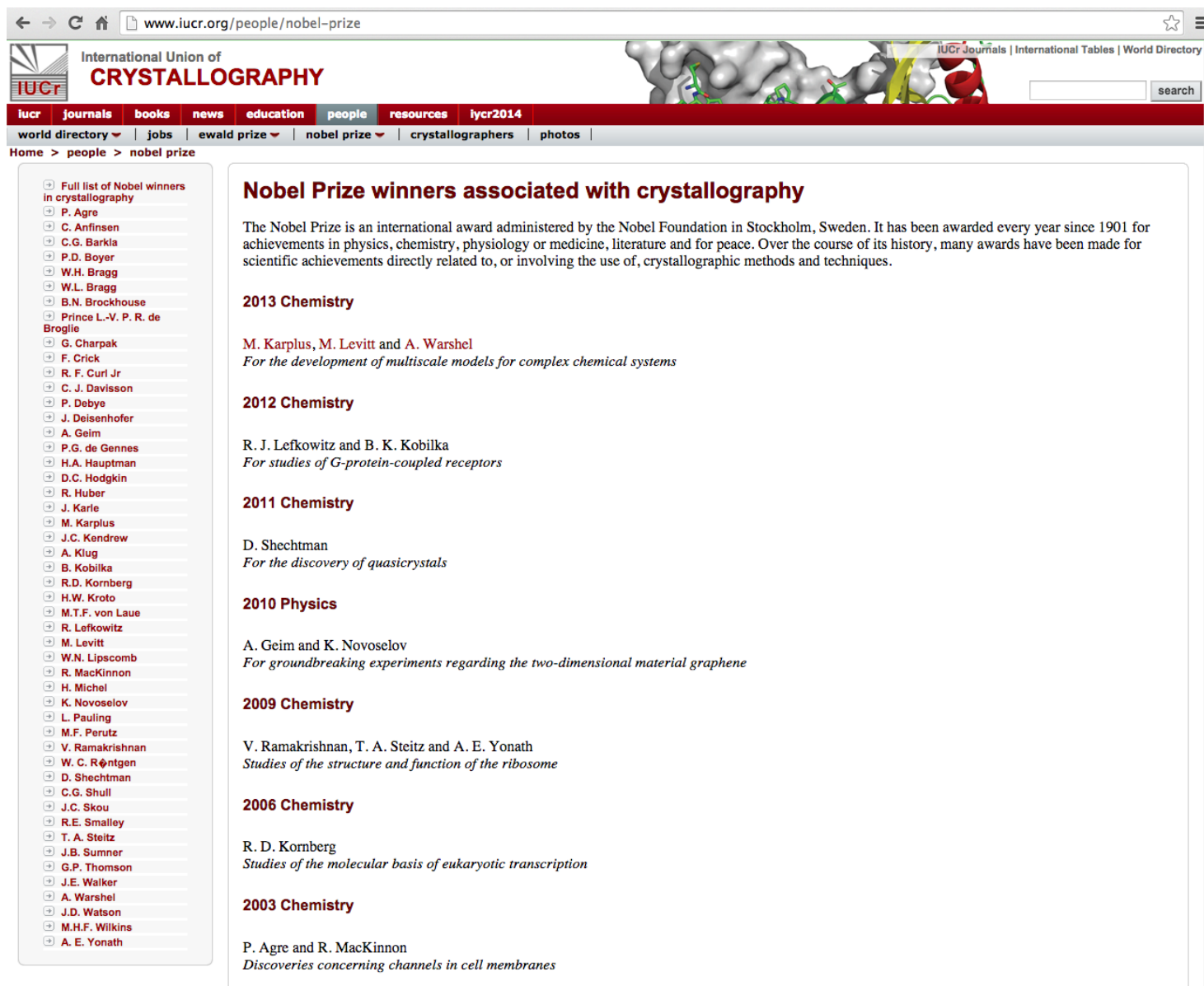
- Protein design (for health or industrial applications)
- Cell design?



How?



A strikingly large share of Nobel Prizes have recognized work on molecular structure



The screenshot shows the IUCr website page for Nobel Prize winners in crystallography. The page features a navigation menu with categories like 'journals', 'books', 'news', 'education', 'people', 'resources', and 'iucr2014'. A search bar is located in the top right corner. The main content area is titled 'Nobel Prize winners associated with crystallography' and provides a detailed list of winners from 2003 to 2013, including their names and the specific work for which they were awarded the Nobel Prize. A sidebar on the left contains a 'Full list of Nobel winners in crystallography' with a scrollable list of names.

International Union of
CRYSTALLOGRAPHY

IUCr Journals | International Tables | World Directory

Home > people > nobel prize

Nobel Prize winners associated with crystallography

The Nobel Prize is an international award administered by the Nobel Foundation in Stockholm, Sweden. It has been awarded every year since 1901 for achievements in physics, chemistry, physiology or medicine, literature and for peace. Over the course of its history, many awards have been made for scientific achievements directly related to, or involving the use of, crystallographic methods and techniques.

2013 Chemistry

M. Karplus, M. Levitt and A. Warshel
For the development of multiscale models for complex chemical systems

2012 Chemistry

R. J. Lefkowitz and B. K. Kobilka
For studies of G-protein-coupled receptors

2011 Chemistry

D. Shechtman
For the discovery of quasicrystals

2010 Physics

A. Geim and K. Novoselov
For groundbreaking experiments regarding the two-dimensional material graphene

2009 Chemistry

V. Ramakrishnan, T. A. Steitz and A. E. Yonath
Studies of the structure and function of the ribosome

2006 Chemistry

R. D. Kornberg
Studies of the molecular basis of eukaryotic transcription

2003 Chemistry

P. Agre and R. MacKinnon
Discoveries concerning channels in cell membranes

Full list of Nobel winners in crystallography

- P. Agre
- C. Anfinsen
- C.G. Barkla
- P.D. Boyer
- W.H. Bragg
- W.L. Bragg
- B.N. Brockhouse
- Prince L.-V. P. R. de Broglie
- G. Charpak
- F. Crick
- R. F. Curl Jr
- C. J. Davison
- P. Dabrye
- J. Deisenhofer
- A. Geim
- P.G. de Gennes
- H.A. Hauptman
- D.C. Hodgkin
- R. Huber
- J. Karle
- M. Karplus
- J.C. Kendrew
- A. Klug
- B. Kobilka
- R.D. Kornberg
- H.W. Kroto
- M.T.F. von Laue
- R. Lefkowitz
- M. Levitt
- W.N. Lipscomb
- R. MacKinnon
- H. Michel
- K. Novoselov
- L. Pauling
- M.F. Perutz
- V. Ramakrishnan
- W. C. Röntgen
- D. Shechtman
- C.G. Shull
- J.C. Skou
- R.E. Smalley
- T. A. Steitz
- J.B. Sumner
- G.P. Thomson
- J.E. Walker
- A. Warshel
- J.D. Watson
- M.H.F. Wilkins
- A. E. Yonath

2013 Nobel Prize recognized early developments underlying modern biomolecular computation

The Nobel Prize in Chemistry 2013



Photo: A. Mahmoud

Martin Karplus

Prize share: 1/3



Photo: A. Mahmoud

Michael Levitt

Prize share: 1/3



Photo: A. Mahmoud

Arieh Warshel

Prize share: 1/3

The Nobel Prize in Chemistry 2013 was awarded jointly to Martin Karplus, Michael Levitt and Arieh Warshel *"for the development of multiscale models for complex chemical systems"*.

2014 Nobel Prize recognized microscopy techniques used to study cellular structure (which also rely on computation)

The Nobel Prize in Chemistry 2014



Photo: A. Mahmoud

Eric Betzig

Prize share: 1/3



Photo: A. Mahmoud

Stefan W. Hell

Prize share: 1/3



Photo: A. Mahmoud

William E. Moerner

Prize share: 1/3

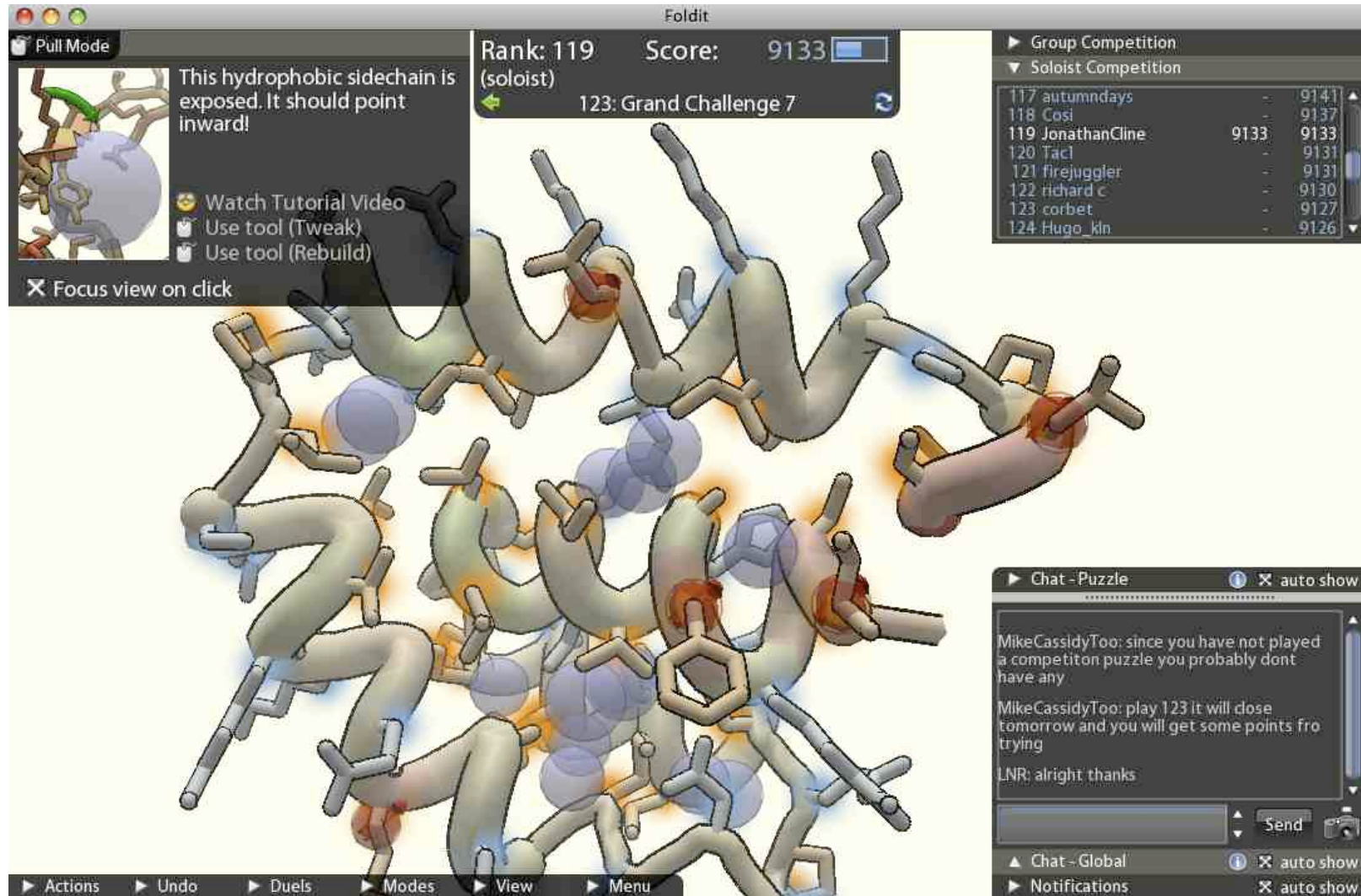
The Nobel Prize in Chemistry 2014 was awarded jointly to Eric Betzig, Stefan W. Hell and William E. Moerner *"for the development of super-resolved fluorescence microscopy"*.

Overview of course topics

Overview of course topics

**Atomic-level modeling of
biomolecules**

Protein structure prediction and RNA/protein design by video game



Foldit - 1-1: One Small Clash

Pull Mode

Score: **7940** of 7900

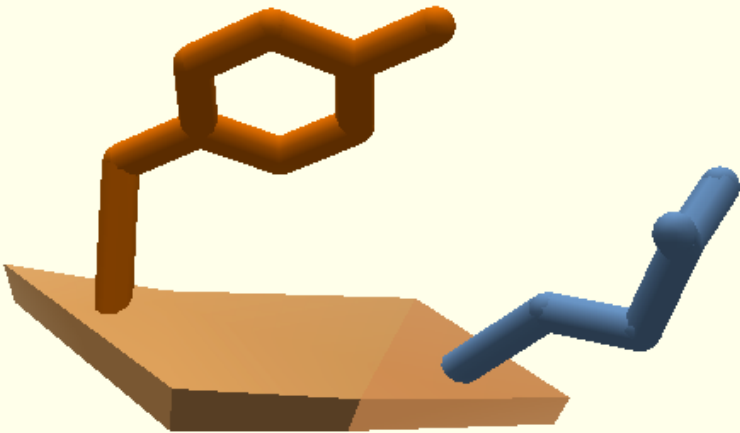
1-1: One Small Clash

You have completed 1 of 31 intro puzzles!

Moves: [★] 4
Time: 0:19

Next is: 1-2: Swing It Around!

 Next Puzzle  Puzzle Menu
 Replay Puzzle [★]

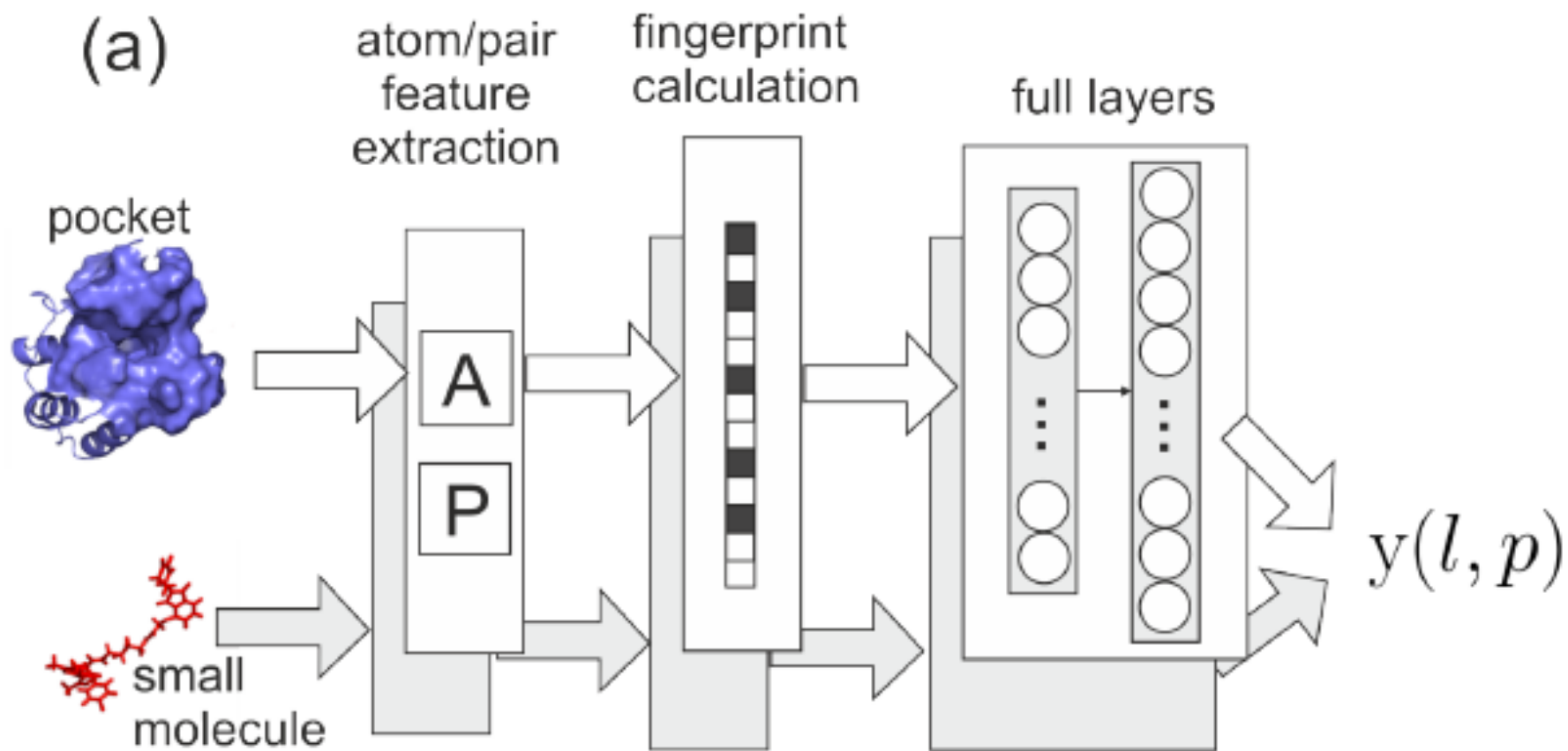


Reset Puzzle

▲ Actions ▶ Undo ▶ Menu

Machine learning on structures

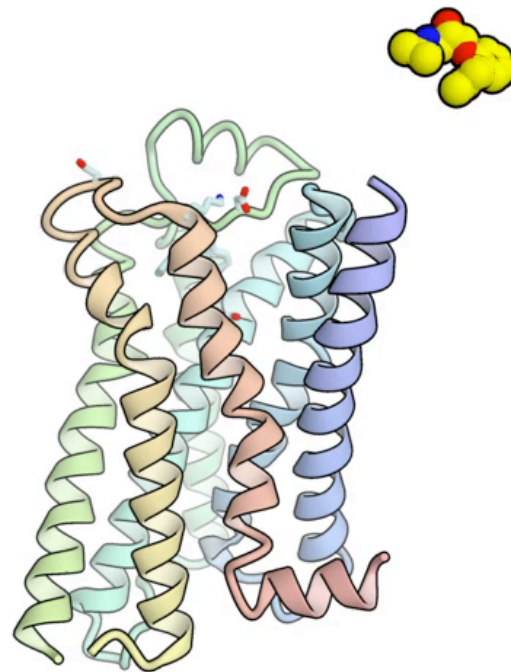
- Applying machine learning techniques to 3D structures of proteins or drug-like molecules — for example, for virtual drug screening



Drug screening: computing accurate binding strengths for drug candidates

- “Alchemical” methods for computing binding free energies: simulate the drug as it gradually “disappears” from the binding pocket or “changes” into a different drug
- Sounds like magic, but it’s actually much more faithful to the physics than traditional docking methods

0.00 us



Coevolution methods for predicting structure from large numbers of genetic sequences

HUMAN	KKASKPKKAASKAPT	KKPKATPVKKAKKK	LAATPKKAKKPKT	VKAKPVKASK
MOUSE	KKAAKPKKAASKAPS	KKPKATPVKKAKKK	PAATPKKAKKPKV	VVKPVKASK
RAT	KKAAKPKKAASKAPS	KKPKATPVKKAKKK	PAATPKKAKKPKI	VKVKPVKASK
COW	KKAAKPKKAASKAPS	KKPKATPVKKAKKK	PAATPKKTKKPKT	VKAKPVKASK
CHIMP	KKASKPKKAASKAPT	KKPKATPVKKAKKK	LAATPKKAKKPKT	VKAKPVKASK

Key idea: amino acids in direct physical contact within a protein tend to mutate in a correlated fashion. Given enough sequence data, one can use this fact to predict structure.

Modern protein design

Design of a protein that selectively transports zinc ions through a cell membrane

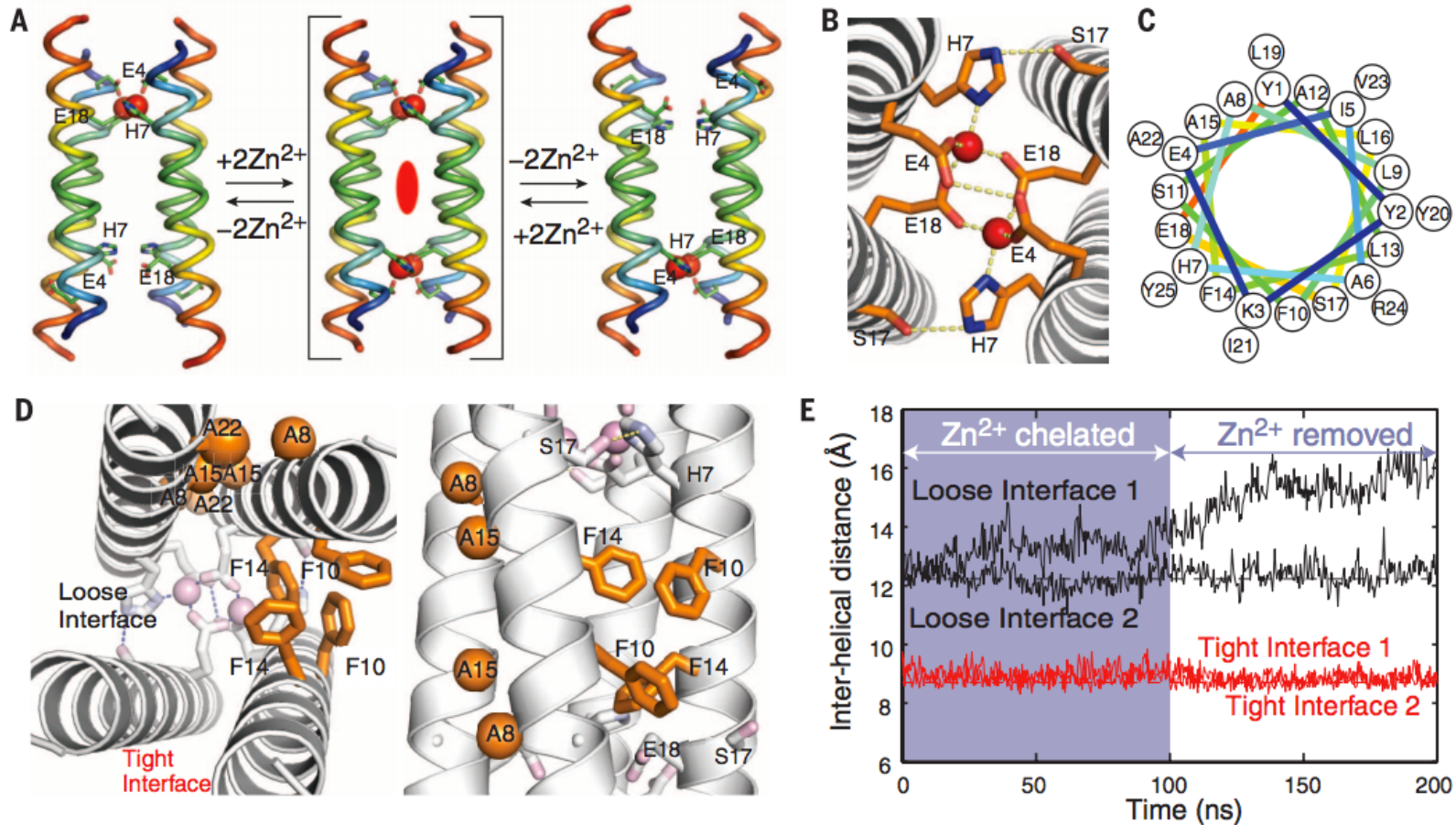
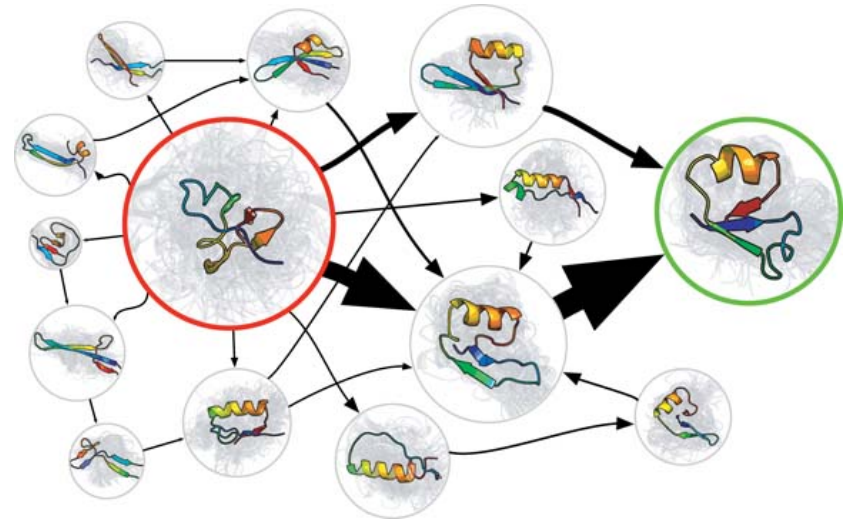
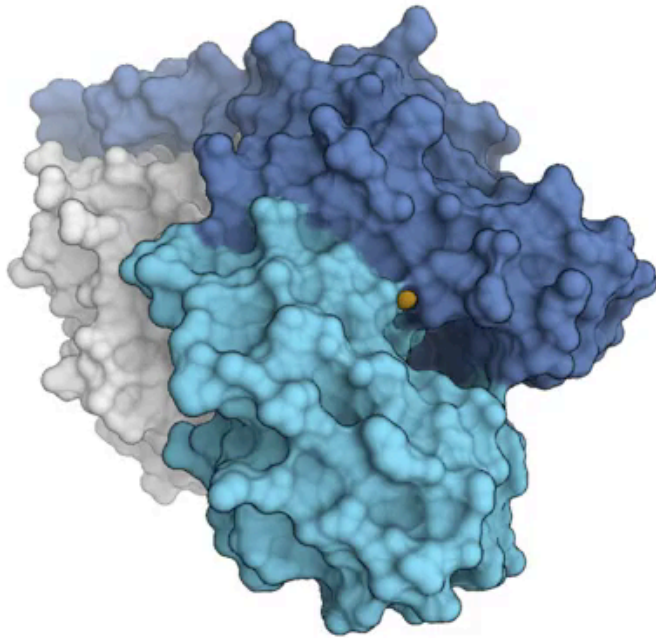


Fig. 1. Computational design and molecular dynamics simulations of Rocker. (A) Schematic of the goal of obtaining conformational exchange between two oppositely oriented symmetry-frustrated states without being trapped in a symmetric state with both sites simultaneously occupied. (B) Metal site consists of a set of ExxH motifs and a single Glu residue from each of the tight dimers. (C) Helical-wheel diagram of Rocker peptide. (D) The re-

packing algorithm placed Ala residues at the tight interface and Phe residues at the loose interface. Empty metal site on the left is omitted for clarity. (E) MD simulation of the design model with two Zn^{2+} ions placed at one metal site show stable interhelical distances for both tight and loose interfaces. Continuing the simulation after removing the Zn^{2+} ions maintained the tight interfaces, but resulted in an increased interhelical distance at the loose interface.

Molecular dynamics simulations and Markov State Models

0.0 us

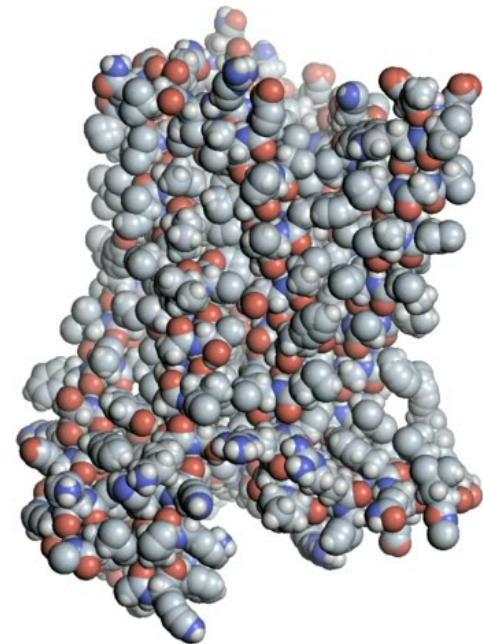
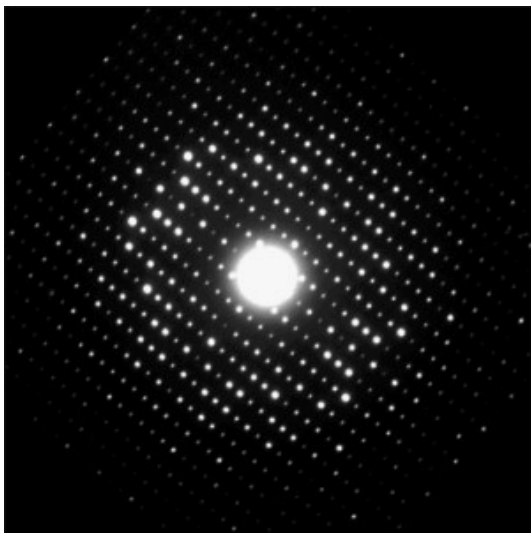


<https://folding.stanford.edu/home/faq/faq-simulation/>

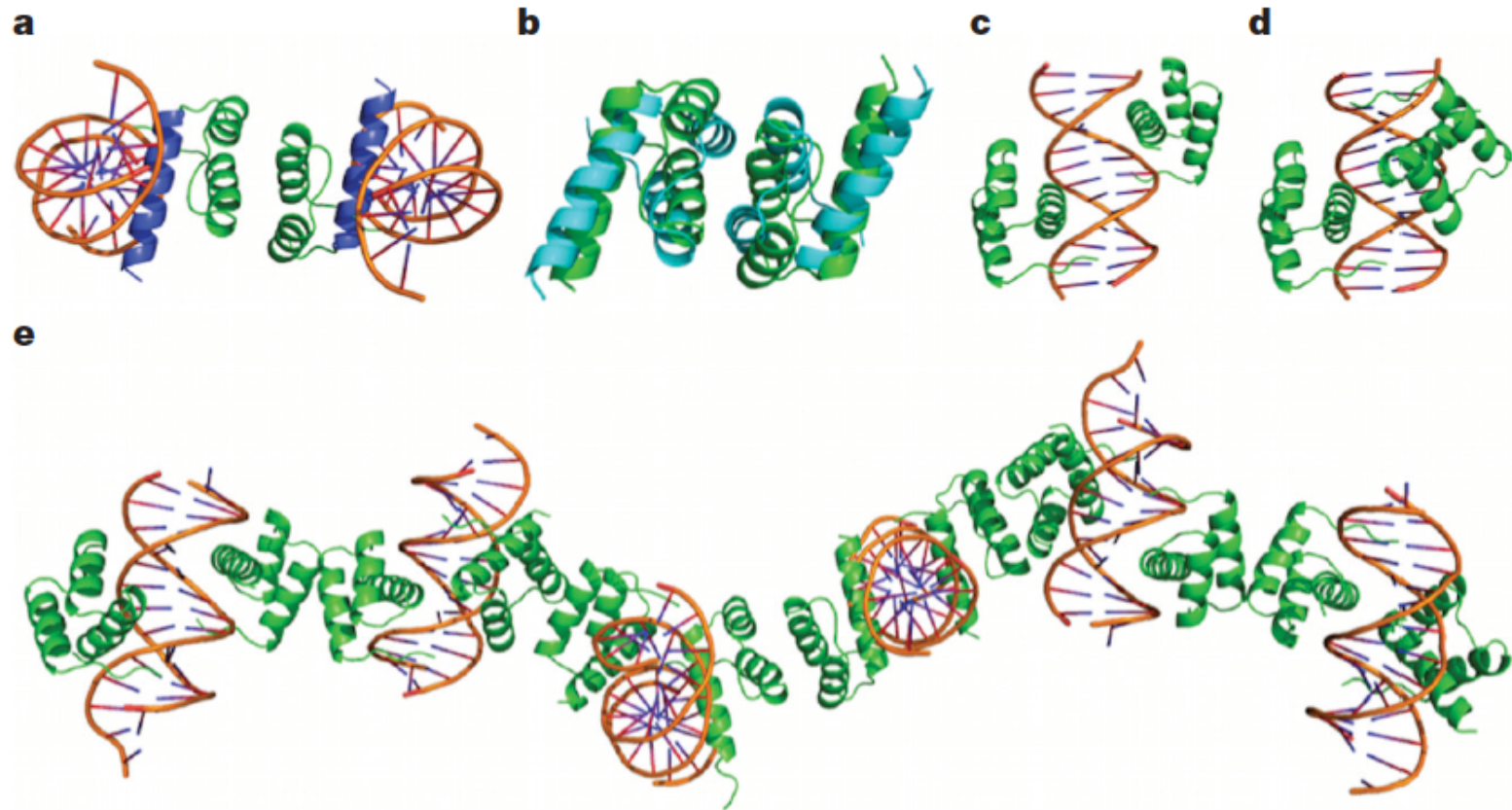
Dror et al., Science 2015

New methods for solving tough crystal structures

- Computational problem in crystallography: go from diffraction pattern to structure
 - Mathematically: invert a Fourier transform without phase information
- New methods for:
 - Solving low-resolution crystal structures using computational protein modeling
 - Solving structures using data from the new x-ray free electron lasers, which vaporize the crystal as they image it



Nucleic acid (i.e., DNA and RNA) structure and design



Computational design of co-assembling protein–DNA nanowires
(Mou et al., *Nature* 2015)

Overview of course topics

Structures of macromolecular complexes

Single-particle electron microscopy

- The current revolution in structure determination through electron microscopy is due in part to new computational methods — e.g., Bayesian approaches



THE REVOLUTION WILL NOT BE CRYSTALLIZED

MOVE OVER X-RAY CRYSTALLOGRAPHY. CRYO-ELECTRON MICROSCOPY IS KICKING UP A STORM IN STRUCTURAL BIOLOGY BY REVEALING THE HIDDEN MACHINERY OF THE CELL.

BY EWEN CALLAWAY

In a basement room, deep in the bowels of a steel-clad building in Cambridge, a major insurgency is under way.

A hulking metal box, some three metres tall, is quietly beaming terabytes' worth of data through thick orange cables that disappear off through the ceiling. It is one of the world's most advanced cryo-electron microscopes: a device that uses electron beams to photograph frozen biological molecules and lay bare their molecular shapes. The microscope is so sensitive that a shout can ruin an experiment, says Sjors Scheres, a structural biologist at the UK Medical Research Council Laboratory of Molecular Biology (LMB), as he stands dwarfed beside the £5-million (US\$7.7-million) piece of equipment. "The UK needs many more of these, because there's going to be a boom," he predicts.

In labs around the world, cryo-electron microscopes such as this one are sending tremors through the field of structural biology. In the past three years, they have revealed exquisite details of protein-making ribosomes, quivering membrane proteins and other key cell molecules.

ILLUSTRATION BY MICHAEL ZUCKER

172 | NATURE | VOL 525 | 10 SEPTEMBER 2015
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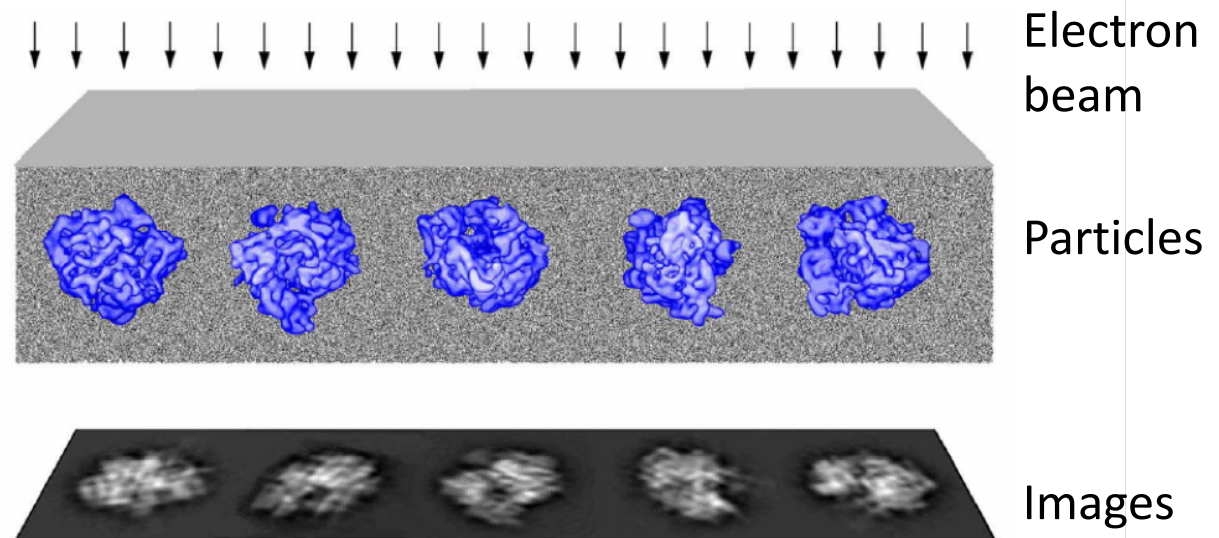


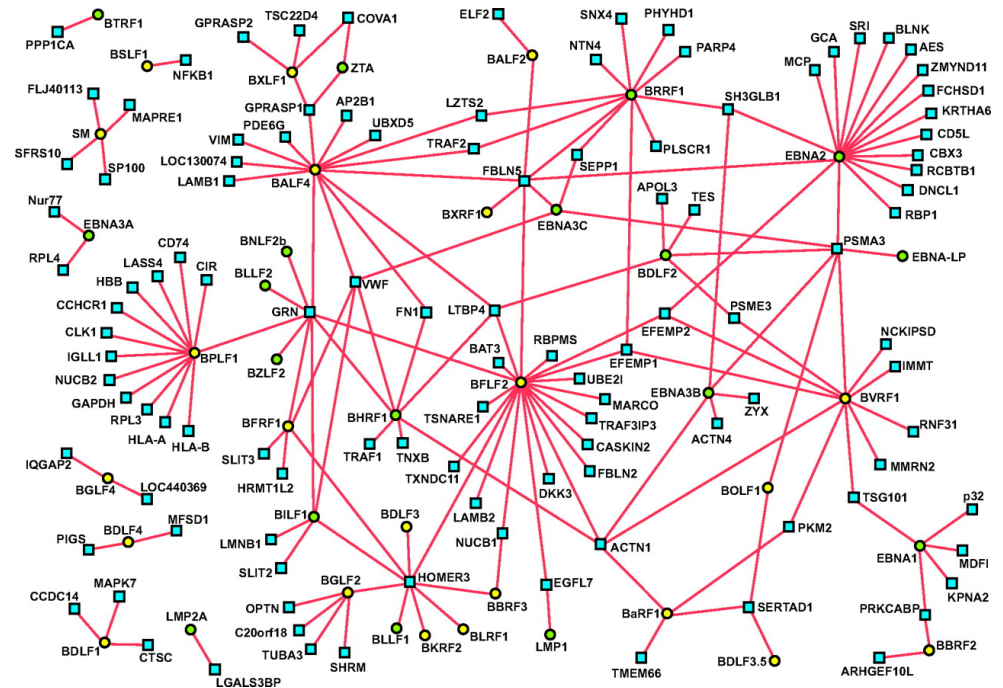
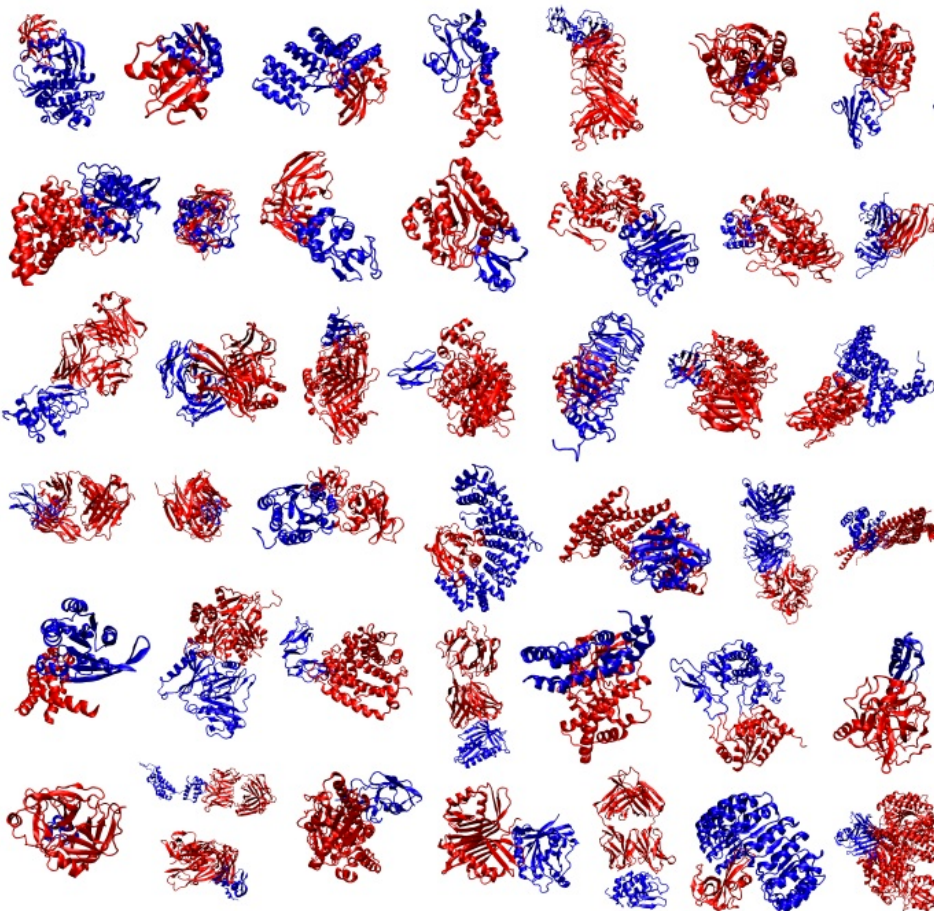
Image from Joachim Frank

<http://biomachina.org/courses/structures/091.pdf>

Nature, Sept. 10, 2015

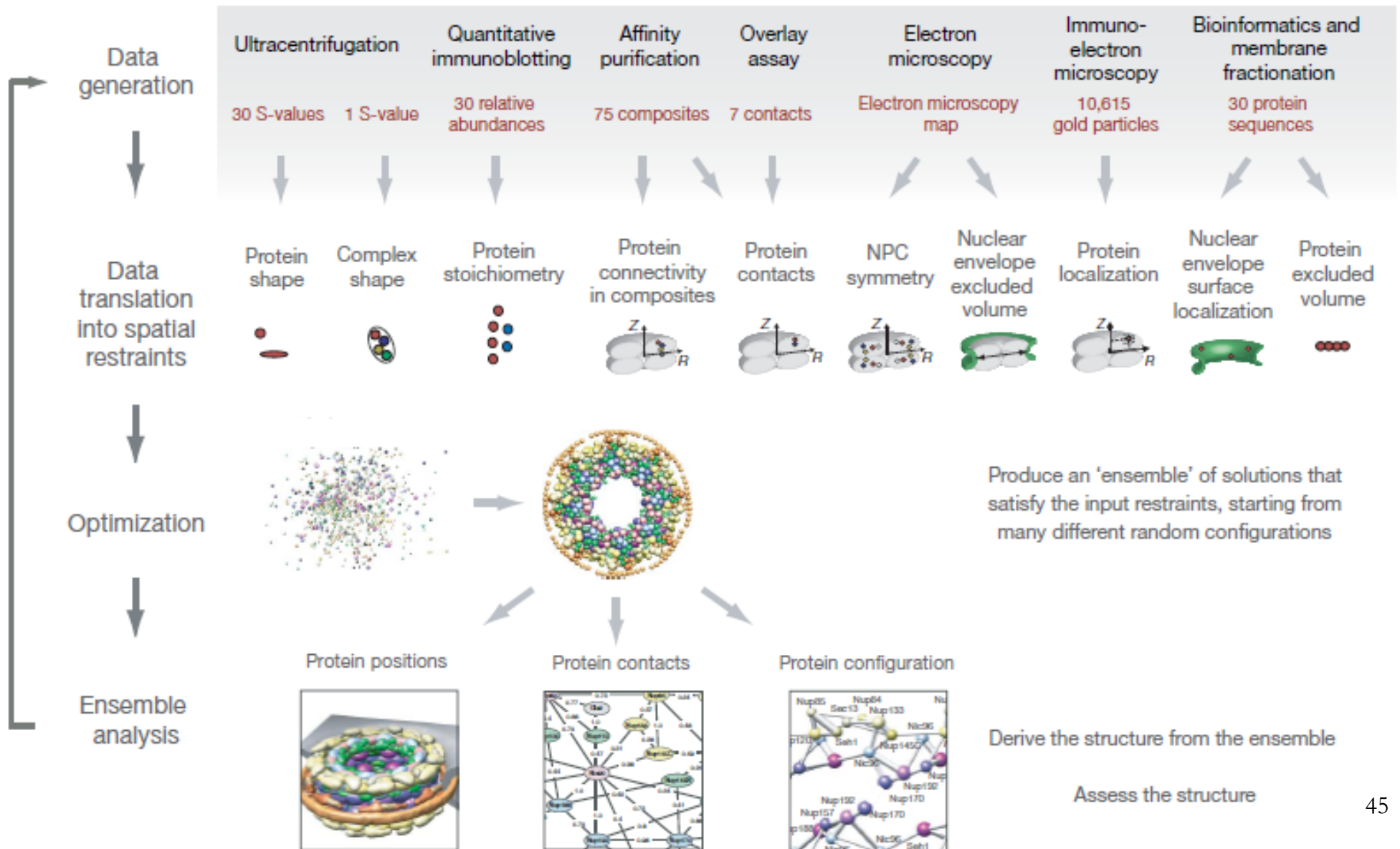
Predicting protein–protein interactions

- Which proteins bind to one another?
- What is the structure of the bound complex?
- What types of networks do they form?



Calderwood et al., PNAS 104:7606-11, 2007

Integrative modeling: combining diverse experimental data to deduce structures of large complexes



Genome architecture

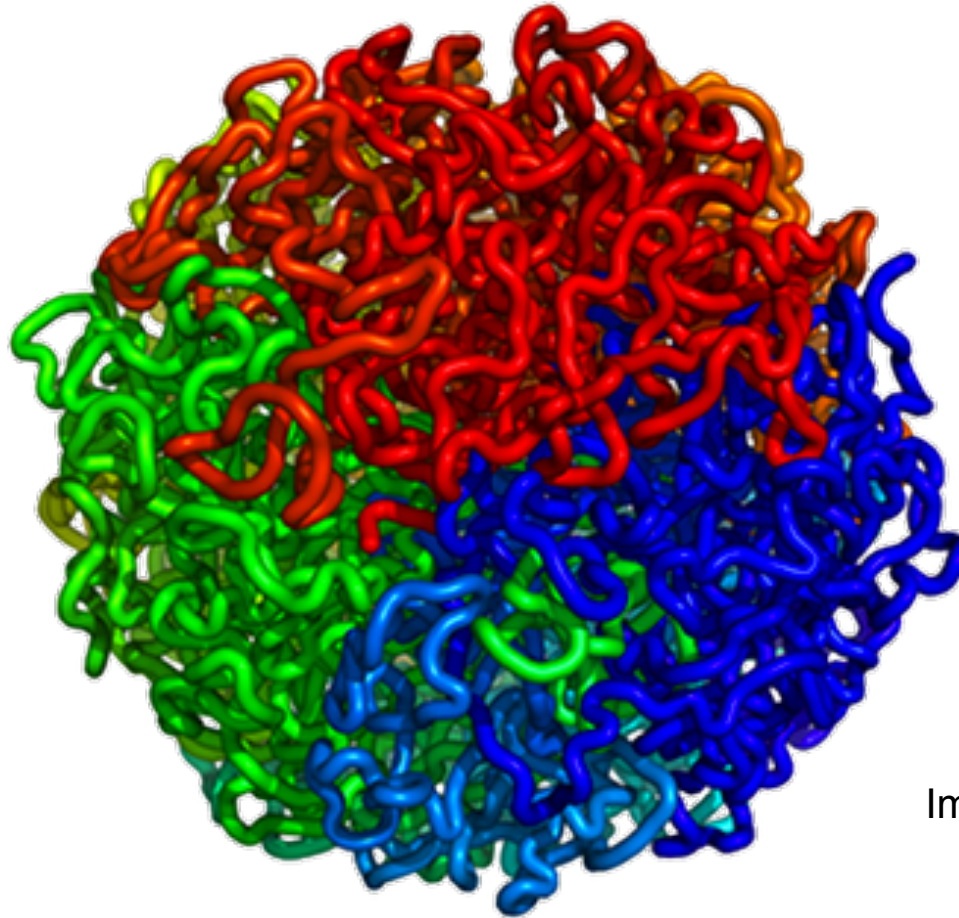


Image: <http://www.aidenlab.org/>

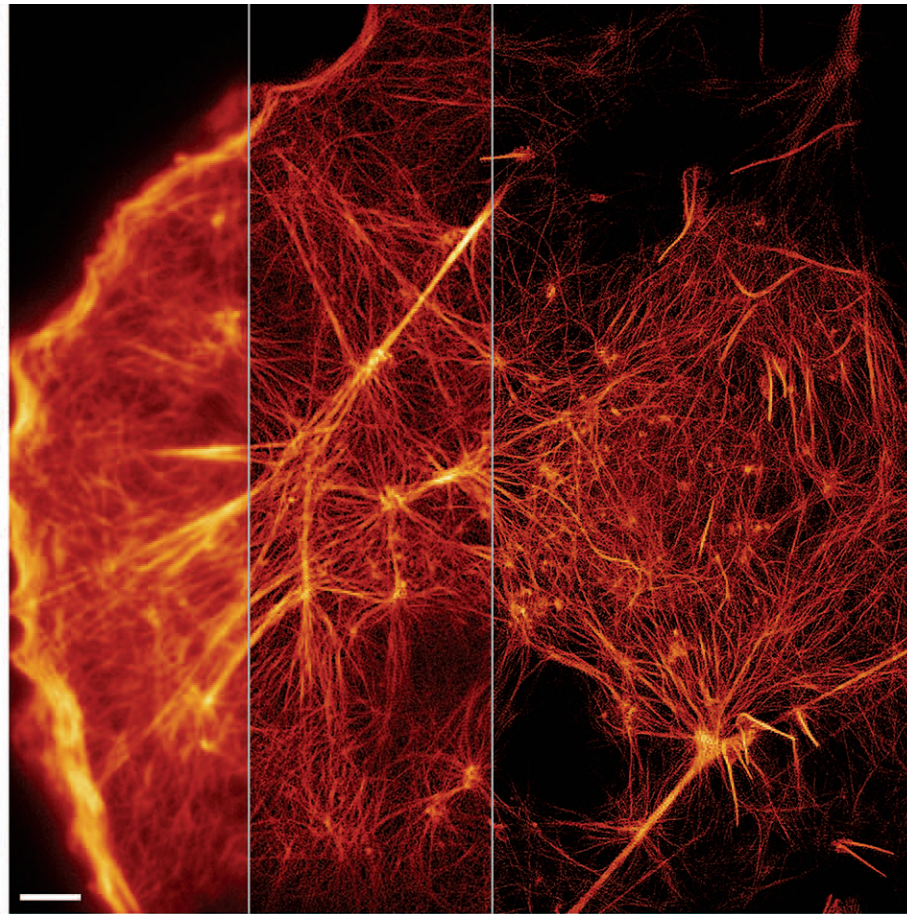
Techniques for mapping out the three-dimensional organization (and dynamics) of your DNA using a combination of computation and experiment

Overview of course topics

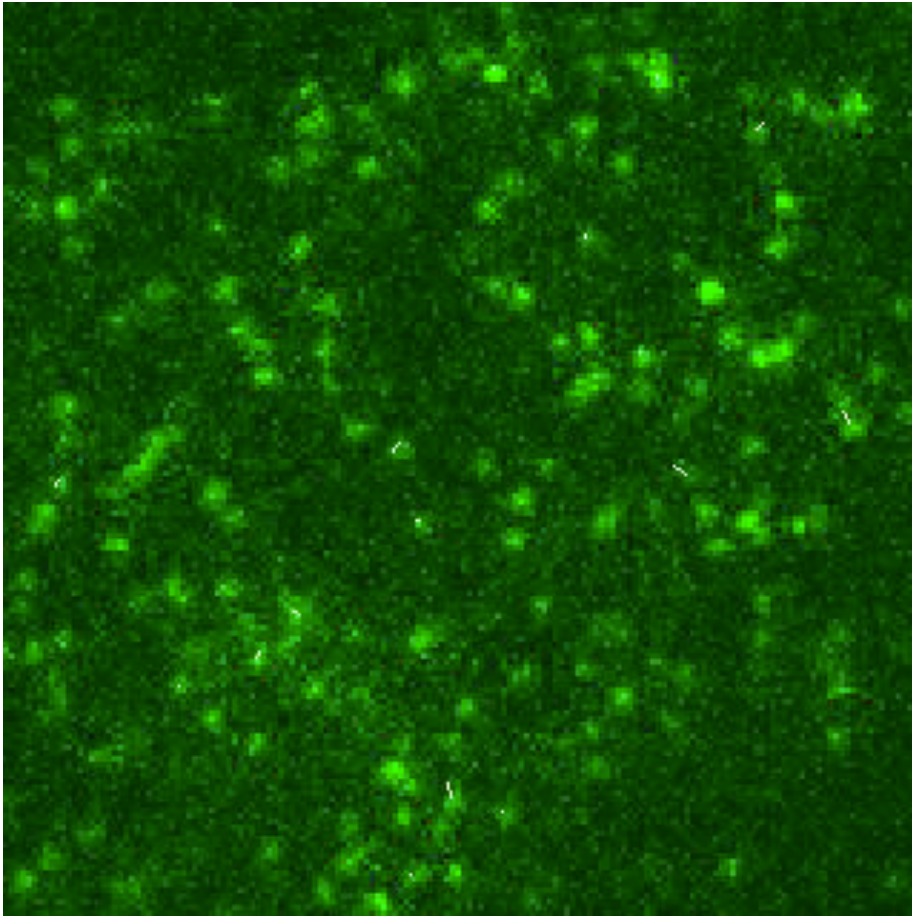
Cellular-level organization

Super-resolution microscopy methods

- New super-resolution microscopy methods
- Analysis methods that exploit compressed sensing



Tracking the motion of single molecules



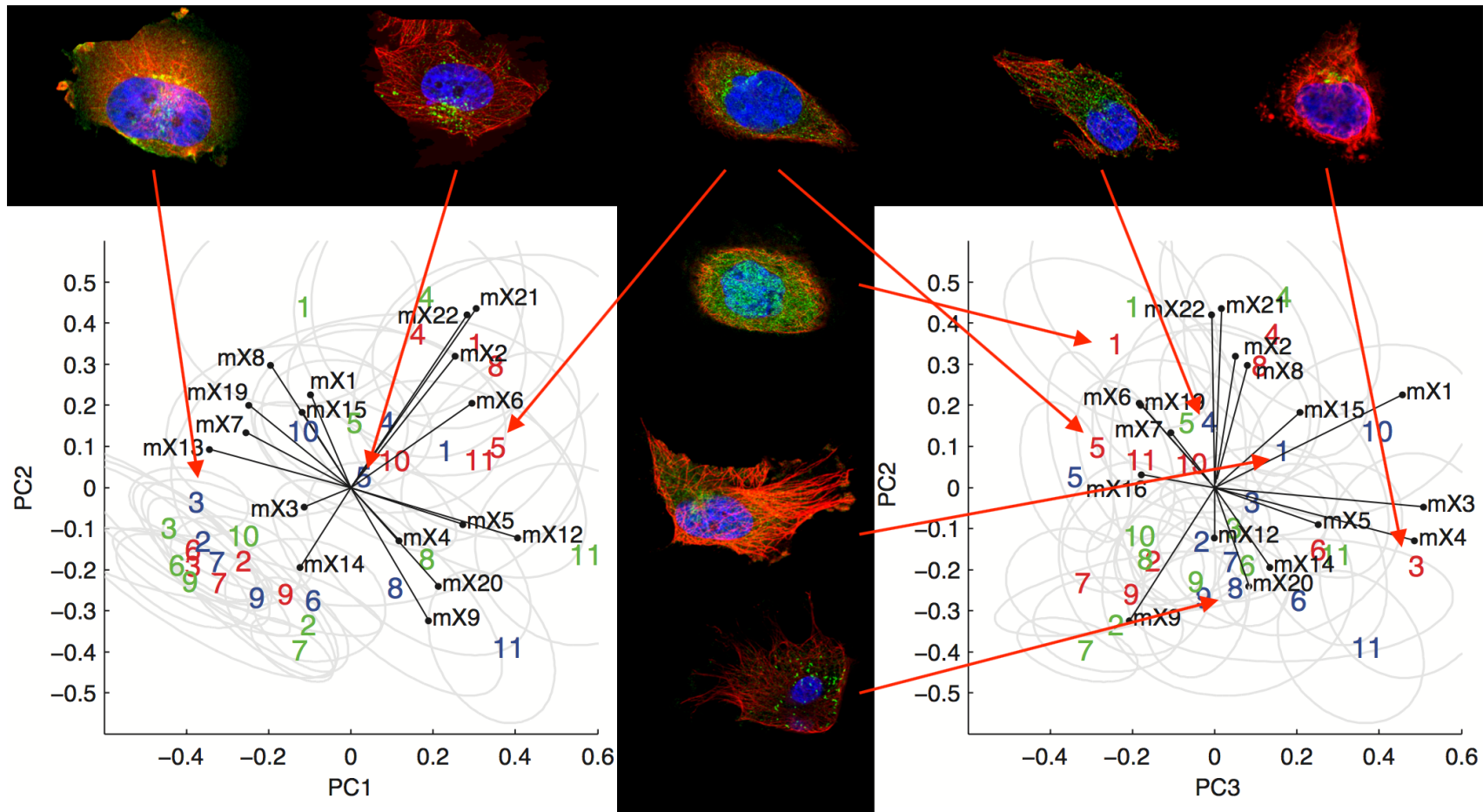
Data: Bettina van Lengerich, Natalia Jura
Tracking and movie: Robin Jia

- Fluorescence microscopy makes it possible to follow the motion of individual molecules in live cells, but actually doing the tracking is a challenging image analysis/computer vision problem

Cellular-level simulations



Learning on cell shape and structure



Course logistics

Course web page

- <http://cs371.stanford.edu/>
- Evaluation criteria on web page
- **Please sign up on Piazza (via link on webpage) so that you get announcements**

Course staff

- Prof. Ron Dror
 - <http://drorlab.stanford.edu/rondror.html>
 - Office hours: Monday and Wednesday after class (2:50-3:45), in classroom and then in Gates 204, or by appointment
- TA: Anthony Ma
 - Office hours: Tuesdays 1:30-2:30, Lathrop Tech Lounge
- TA: Osama El-Gabalawy
 - Office hours TBA

Expected background

- Course is intended to be broadly accessible to students with computational or biological backgrounds
- You should have:
 - Introductory biology background, at the level of a first-year course, and ideally also introductory chemistry and physics background
 - Introductory computer science background (e.g., CS 106A)
 - Math through calculus, and some previous exposure to mathematical modeling or probability.
- You're *not* expected to be an expert in any of the areas we cover, but you *are* expected to be willing to read and present papers, ask questions, and share your thoughts

Honor code

- I'm not a tough grader, but I take the honor code seriously
- In the context of this course, this means:
 - Avoid verbatim **or near-verbatim** copying of text from any source (unless it's enclosed in quotation marks and clearly attributed to the source). You need to explain ideas in your own words.
 - Make sure that all materials (including figures) in presentations and critiques are properly cited.

Guidelines for presentations and critiques

Giving a good presentation

- Why should you care?
 - To develop a critical skill
 - To help make this a good course!
- See “Tips for Giving Clear Talks” by Kayvon Fatahalian (Stanford CS PhD 2010)
 - <http://www.cs.cmu.edu/~kayvonf/misc/cleartalktips.pdf>
 - This focuses on giving talks on your own research but is applicable to the presentations for this class as well. Possible exception: I'd like you to point out both strengths and weaknesses in the work you present.

Giving a good presentation

- Make it understandable to a diverse audience
 - Students in class are from: CS, CME, EE, BioE, ChemE, chemistry, chemical & systems biology, biophysics, and more
 - Include appropriate background material. You may need to read other, related papers as well.
- Describe the main idea *intuitively*
 - *You need to figure out what the main idea is.* (Ask course staff for help if necessary.)
 - Do not rely on equations or code to convey intuition
 - You do not need to cover everything in the paper
- Strive for clarity
 - See Kayvon's presentation
- Make it interesting and exciting!

Preparing for presentations

- Each presenter should meet with a TA, and then with me (Ron), before their presentation
 - By default, meet with me right after the class preceding your presentation. If one of the presenters has a conflict then, schedule a different time.
- You should prepare a complete draft of your presentation before these meetings
- You need to coordinate with the other students presenting on the same day.
 - For example, you may want shared introductory material
 - I would like to meet with all of you together

Writing a critique

- Although you should include a brief summary of the paper you're critiquing, that shouldn't be the focus
- Instead, the focus should be on:
 - Strengths and weaknesses of the approach or results
 - Other approaches that might have been applied instead
 - Potential extensions or follow-on work
- You're encouraged to read related papers to help you write this.
 - Please indicate in the critique which other papers you read.

Immediate next steps

Choosing topics for presentation and critique

- By next Tuesday, Jan. 17, email cs371-win1617-staff@lists.stanford.edu a list of six topics (class sessions) you'd be interested in presenting on and twelve papers (or six topics) you'd be interested in critiquing
 - My default list of papers is at <http://cs371.stanford.edu/schedule.html> and will be complete by Tuesday afternoon. The “Main Papers” are the ones I’m expecting students to present.
 - You’re also welcome to suggest your own paper(s) of interest to present. If you do, specify any dates on which you can’t present.
 - You may rank order your lists if you’d like (you should specify that you’ve done so)
- Also include:
 - Brief description of your background

Next couple class periods

- Wednesday, Jan. 11: Ron to present on simulation of drug targets and simulation analysis
 - Reading materials on website
- Monday, Jan. 16: Guest lecture by Eli Groban (Autodesk) on virtual reality for biomolecules
 - *Completely optional (MLK day).*

Volunteers for initial presentations & critiques

- Email cs371-win1617-staff@lists.stanford.edu ASAP to volunteer to present or critique during the following four class periods:
 - Jan. 18: Using multiplayer online video games for structure prediction and design
 - Jan. 23: Coevolution methods for predicting structure from large numbers of genetic sequences
 - Jan. 25: Modern protein design
 - Jan. 27: Machine learning for structure-based virtual screening